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<b>(54) Title:</b> CHIMERIC ADENOVIRAL VECTORS  <b>(57) Abstract</b>  A chimeric adenoviral vector is provided that comprises nucleotide sequence of a first adenovirus, wherein all or part of at least one gene of said first adenovirus encoding a protein that facilitates binding of said vector to a target mammalian cell, or internalization thereof within said cell, is replaced by all or part of the corresponding gene from a second adenovirus belonging to subgroup D, said vector further comprising a transgene operably linked to a eucaryotic promoter to allow for expression therefrom in a mammalian cell. Compositions comprising such vectors and methods of using such vectors to deliver transgenes to target mammalian cells, particularly airway epithelial cells, are also provided.		

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Description

## Chimeric Adenoviral Vectors

5. Introduction

The present invention relates to chimeric adenoviral vectors, that is, vectors comprising DNA from more than one serotype of adenovirus, which offer enhanced infection efficiency of target cells in order to deliver one or more therapeutically useful nucleotide sequences, including transgenes, therein. Such a nucleotide sequence may comprise a gene not otherwise present in the target cell that codes for a therapeutic and/or biologically active protein, or may represent, for example, an active copy of a gene that is already present in the target cell, but in a defective or deficient form.

15. Background of the Invention

One of the fundamental challenges now facing medical practitioners is that although the defective genes that are associated with numerous inherited diseases (or that represent disease risk factors including for various cancers) have been isolated and characterized, methods to correct the disease states themselves by providing patients with normal copies of such genes (the technique of gene therapy) are substantially lacking. Accordingly, the development of improved methods of intracellular delivery therefor is of great medical importance. Examples of diseases that it is hoped can be treated by gene therapy include inherited disorders such as cystic fibrosis, Gaucher's disease, Fabry's disease, and muscular dystrophy.

25 Representative of acquired disorders that can be treated are: (1) for cancers: multiple myeloma, leukemias, melanomas, ovarian carcinoma and small cell lung cancer; (2) for cardiovascular conditions: progressive heart failure, restenosis, and hemophilias; and (3) for neurological conditions: traumatic brain injury.

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Gene therapy requires successful transfer of nucleic acid to the target cells of a patient. Gene transfer may generally be defined as the process of introducing an expressible polynucleotide (for example a gene, a cDNA, or an mRNA patterned thereon) into a cell. In a particular application of this approach, successful expression  
5 of an encoding polynucleotide leads to production in the cells of a normal protein and leads to correction of a disease state associated with an abnormal gene. Therapies based on providing such proteins directly to target cells (protein replacement therapy) have generally proved ineffective since, for example, the cell membrane presents a selectively permeable barrier to entry. Thus there is great interest in alternative  
10 methods to cause delivery of therapeutic proteins, especially by transfer of the relevant polynucleotide, often referred to as a transgene.

Viral vectors have been used with increasing frequency to date to deliver transgenes to target cells. Most attempts to use viral vectors for gene therapy have relied on retrovirus-based vectors, chiefly because of their ability to integrate into the  
15 cellular genome. However, the disadvantages of retroviral vectors are becoming increasingly clear, including their tropism for dividing cells only, the possibility of insertional mutagenesis upon integration into the cell genome, decreased expression of the transgene over time, rapid inactivation by serum complement, and the possibility of generation of replication-competent retroviruses. See, for example, D. Jolly, et al.,  
20 Cancer Gene Therapy, 1, 1994, pp. 51-64, and C.P. Hodgson, et al., Bio Technology , 13, 1995, pp. 222-225. Such disadvantages have led to the development of other viral-based vector systems, including those derived from adenoviruses.

Adenovirus (Ad) is a nuclear DNA virus with a genome of about 36 kb, which has been well-characterized through studies in classical genetics and molecular  
25 biology. A detailed discussion of adenovirus is found in Thomas Shenk, "Adenoviridae and their Replication", and M. S. Horwitz, "Adenoviruses" , Chapters 67 and 68, respectively, in Virology, B.N. Fields et al., eds., 2nd edition, Raven Press, Ltd., New York, 1996, and reference therein is found to numerous aspects of adenovirus pathology, epidemiology, structure, replication, genetics and classification.

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In a simplified form, the adenoviral genome is classified into early (known as E1-E4) and late (known as L1-L5) transcriptional units, referring to the generation of two temporal classes of viral proteins. The demarcation between these events is viral DNA replication.

5       The human adenoviruses are divided into numerous serotypes (approximately 47, numbered accordingly and classified into 6 subgroups: A, B, C, D, E and F), based upon properties including hemagglutination of red blood cells, oncogenicity, DNA base and protein amino acid compositions and homologies, and antigenic relationships. Additional background information concerning Ad serotype  
10 classification, including that for subgroup D, can be found, for example, in F. Deryckere et al., *Journal of Virology*, 70, 1996, pp. 2832-2841; and A. Bailey et al., *Virology*, 205, 1994, pp. 438-452, and in other art-recognized references.

Adenoviruses are nonenveloped, regular icosahedrons (having 20 triangular surfaces and 12 vertices) that are about 65-80 nm in diameter. A protein called fiber  
15 projects from each of these vertices. The fiber protein is itself generally composed of 3 identical polypeptide chains, although the length thereof varies between serotypes. The protein coat (capsid) is composed of 252 subunits (capsomeres), of which 240 are hexons, and 12 are pentons. Each penton comprises a penton base, on the surface of the capsid, and a fiber protein projecting from the base. The Ad 2 penton base protein,  
20 for example, has been determined to be a 8 x 9 nm ring shaped complex composed of 5 identical protein subunits of 571 amino acids each.

Current understanding of adenovirus-cell interactions suggests that adenovirus utilizes two cellular receptors to attach to, and then infect a target cell. It has been further suggested that the fiber protein of an infecting adenovirus first attaches to a  
25 receptor, the identity of which is still unknown, and then penton base attaches to a further receptor, often a protein of the alpha integrin family. It has been determined that alpha-integrins often recognize short amino acid sequences on other cellular proteins for attachment purposes including the tripeptide sequence Arg-Gly-Asp (abbreviated RGD). An RGD sequence is also found in the penton base protein of

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adenovirus and is currently understood in the art to mediate attachment of Ad to alpha integrins.

Recombinant adenoviruses have several advantages for use as gene transfer vectors, including tropism for both dividing and non-dividing cells, minimal pathogenic potential, ability to replicate to high titer for preparation of vector stocks, and the potential to carry large inserts (Berkner, K.L., *Curr. Top. Micro. Immunol.* 158:39-66, 1992; Jolly, D., *Cancer Gene Therapy* 1:51-64, 1994).

The carrying capacity of an adenovirus vector is proportional to the size of the adenovirus genome present in the vector. For example, a capacity of about 8 kb can be created from the deletion of certain regions of the virus genome dispensable for virus growth, e.g., E3, and the deletion of a genomic region such as E1 whose function may be restored in trans from 293 cells (Graham, F.L., *J. Gen. Virol.* 36:59-72, 1977) or A549 cells (Imler et al., *Gene Therapy* 3:75-84, 1996). Such E1-deleted vectors are rendered replication-defective, which is desirable for the engineering of adenoviruses for gene transfer. The upper limit of vector DNA capacity for optimal carrying capacity is about 105%-108% of the length of the wild-type genome. Further adenovirus genomic modifications are possible in vector design using cell lines which supply other viral gene products in trans, e.g., complementation of E2a (Zhou et al., *J. Virol.* 70:7030-7038, 1996), complementation of E4 (Krougliak et al., *Hum. Gene Ther.* 6:1575-1586, 1995; Wang et al., *Gene Ther.* 2:775-783, 1995), or complementation of protein IX (Caravokyri et al., *J. Virol.* 69:6627-6633, 1995; Krougliak et al., *Hum. Gene Ther.* 6:1575-1586, 1995). Maximal carrying capacity can be achieved using adenoviral vectors deleted for all viral coding sequences (Kochanek et al., *Proc. Natl. Acad. Sci. USA* 93:5731-5736, 1996; Fisher et al., *Virology* 217:11-22, 1996).

Transgenes that have been expressed to date by adenoviral vectors include p53 (Wills et al., *Human Gene Therapy* 5:1079-188, 1994); dystrophin (Vincent et al., *Nature Genetics* 5:130-134, 1993; erythropoietin (Descamps et al., *Human Gene Therapy* 5:979-985, 1994; ornithine transcarbamylase (Stratford-Perricaudet et al.,

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Human Gene Therapy 1:241-256, 1990; We et al., J. Biol. Chem. 271:3639-3646, 1996;); adenosine deaminase (Mitani et al., Human Gene Therapy 5:941-948, 1994); interleukin-2 (Haddada et al., Human Gene Therapy 4:703-711, 1993); and  $\alpha$ 1-antitrypsin (Jaffe et al., Nature Genetics 1:372-378, 1992); thrombopoietin  
5 (Ohwada et al., Blood 88:778-784, 1996); and cytosine deaminase (Ohwada et al., Hum. Gene Ther. 7:1567-1576, 1996).

The particular tropism of adenoviruses for cells of the respiratory tract has particular relevance to the use of adenovirus in gene therapy for cystic fibrosis (CF), which is the most common autosomal recessive disease in Caucasians. The disease is  
10 caused by the presence of one or more mutations in the gene that encodes a protein known as cystic fibrosis transmembrane conductance regulator (CFTR), and which regulates the movement of ions (and therefore fluid) across the cell membrane of epithelial cells, including lung epithelial cells. Abnormal ion transport in airway cells leads to abnormal mucous secretion, inflammation and infection, tissue damage,  
15 and eventually death. Mutations in the CFTR gene that disturb the cAMP-regulated  $\text{Cl}^-$  channel in airway epithelia result in pulmonary dysfunction (Zabner et al., Nature Genetics 6:75-83, 1994). Adenovirus vectors engineered to carry the CFTR gene have been developed (Rich et al., Human Gene Therapy 4:461-476, 1993) and studies have shown the ability of these vectors to deliver CFTR to nasal epithelia of CF patients  
20 (Zabner et al., Cell 75:207-216, 1993), the airway epithelia of cotton rats and primates (Zabner et al., Nature Genetics 6:75-83, 1994), and the respiratory epithelium of CF patients (Crystal et al., Nature Genetics 8:42-51, 1994). Recent studies have shown that administering an adenoviral vector containing a DNA sequence encoding CFTR to airway epithelial cells of CF patients can restore a functioning chloride ion channel  
25 in the treated epithelial cells (Zabner et al., J. Clin. Invest. 97:1504-1511, 1996; U.S. Patent No. 5,670,488 issued September 23, 1997).

Serotype classification is partly based on viral surface protein sequence variation. Because the infectious capabilities of the virus are associated with the surface protein interactions of the virus with cellular proteins, the serotype is an



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important determinant of viral entry into target cells, and can account for the infectious heterogeneity of adenovirus serotypes. Most adenoviral vectors have been constructed using adenovirus serotypes from the well-studied group C adenoviruses, especially Ad 2 and Ad 5. However, other adenovirus serotypes display infectious  
5 properties that are relevant to the further design of improved adenoviral vectors, for example, those derived from subgroup D, which display enhanced tropism for human airway epithelial cells.

It is widely hoped that gene therapy will provide a long lasting and predictable form of therapy for certain disease states, and it is likely the only form of therapy  
10 suitable for many inherited diseases. Although adenoviral vectors are currently in clinical use and have shown therapeutic promise, a need remains to improve the infection efficiency of these vectors in order to further improve their gene transfer capabilities. The present invention addresses this goal.

### 15 Summary Of The Invention

The present invention provides for chimeric adenoviral vectors which offer enhanced infection efficiency of target cells for the delivery of one or more transgenes. In a representative aspect of the invention, the vectors comprise nucleotide sequences coding for therapeutically useful proteins and have enhanced tropism for airway  
20 epithelial cells.

Accordingly, there are provided chimeric adenoviral vectors comprising nucleotide sequence of a first adenovirus, wherein at least one gene of said first adenovirus encoding a protein that facilitates binding of said vector to a target mammalian cell, or internalization thereof within said cell, is replaced by the  
25 corresponding gene from a second adenovirus belonging to subgroup D. These vectors may further comprising a transgene operably linked to a eucaryotic promoter or other regulatory elements to allow for expression therefrom in a mammalian cell. In a representative aspect thereof, the replaced encoding sequence codes for Ad fiber, hexon or penton base.



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In a further preferred embodiment of the invention, there are provided chimeric adenoviral vectors comprising nucleotide sequence of a first adenovirus, wherein a portion of a gene thereof encoding a protein that facilitates binding of said vector to a target mammalian cell, or internalization thereof within said cell, is replaced by a  
5 portion of the corresponding gene from a second adenovirus belonging to subgroup D. These vectors may further comprising a transgene operably linked to a eucaryotic promoter or other regulatory elements to allow for expression therefrom in a mammalian cell. In a representative aspect thereof, the replaced encoding sequence codes for a portion of Ad fiber, hexon or penton base.

10 Preferably, the second adenovirus is a member of subgroup D, and the replaced nucleotide sequence encodes a polypeptide selected from the group consisting of Ad fiber, a fragment of Ad fiber, Ad hexon, a fragment of Ad hexon, Ad penton base, and a fragment of Ad penton base. In a preferred embodiment, said second adenovirus is selected from the group consisting of serotypes Ad 9, Ad 15, Ad  
15 17, Ad 19, Ad 20, Ad 22, Ad 26, Ad 27, Ad 28, Ad 30, and Ad 39. In preferred embodiments of the chimeric adenoviral vectors, the first adenovirus is selected from the group consisting of Ad 2, Ad 5, and Ad 12.

The invention is also directed to compositions comprising the chimeric adenoviral vectors of the invention. Additional aspects of the invention include  
20 methods to use the chimeric adenoviral vectors of the invention to deliver transgenes to mammalian target cells, for example, to the airway epithelial cells of patients.

A still further representative aspect of the invention involves a method of providing a therapeutic and/or biologically active protein to the airway epithelial cells of a patient by administering to said cells an adenoviral vector comprising elements of  
25 an Ad 17 genome, and a transgene encoding said therapeutic protein that is operably linked to a eucaryotic promoter to allow for expression therefrom in a mammalian cell, under conditions whereby the transgene encoding said therapeutic protein is expressed, and therapeutic benefit is produced in said airway epithelial cells.

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These and other aspects of the present invention are described in the Detailed Description of the Invention which follows directly.

Brief Description of the Drawings

- 5       FIGURE 1 depicts infection of NHBE cells by Ad 2.  
FIGURE 2 depicts infection of NHBE cells by Ad 17.  
FIGURE 3 plots the result of binding to human nasal polyp epithelial cell isolates by Ad 2 and Ad 17.  
FIGURE 4 is a map of the vector Ad2/ $\beta$ gal-2/fiber Ad 17.  
10       FIGURE 5 shows a comparison of the amino acid sequence of penton base from Ad 17 (top) [SEQ ID NO: 4] and Ad 2 (bottom) [SEQ ID NO: 5], and further depicts the variable RGD containing region.  
FIGURE 6 depicts an amino acid sequence pileup for penton base from particular Ad serotypes, including f10 (from fowl) [SEQ ID NO: 6 through SEQ ID  
15   NO: 10].  
FIGURE 7 shows a comparison of the amino acid sequence of fiber from Ad 17 (top) [SEQ ID NO: 11] and Ad 2 (bottom) [SEQ ID NO: 12].  
FIGURE 8 depicts an amino acid sequence pileup for fiber from particular Ad serotypes [SEQ ID NO: 11 through SEQ ID NO: 22], including two forms of serotype  
20   40 (40-1 and 40-2) which differ in that one variant has two (but non-identical) copies of the fiber gene.  
FIGURE 9 shows the infection efficiency of colon cancer cell lines by adenovirus serotypes.  
FIGURE 10 shows the infection efficiency of cancer cell lines by adenovirus  
25   serotypes.  
Provided in the Sequence Listing attached hereto are also:  
SEQ ID NO: 1, the complete nucleotide sequence of Ad 17;  
SEQ ID NO: 2, the complete encoding nucleotide sequence for Ad 17 fiber;

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SEQ ID NO: 3, the complete encoding nucleotide sequence for Ad 17 penton base.

#### Detailed Description of the Invention

5       The present invention provides for chimeric adenoviral vectors comprising nucleotide sequence of a first adenovirus, wherein at least one gene of said first adenovirus encoding a protein that facilitates binding of said vector to a target mammalian cell, or internalization thereof within said cell, is replaced by the corresponding gene from a second adenovirus belonging to subgroup D, said vectors  
10 further comprising a transgene operably linked to a eucaryotic promoter to allow for expression therefrom in a mammalian cell. In a representative aspect thereof, the replaced encoding sequence correspond to the gene encoding the Ad fiber, hexon or penton base proteins, or combinations thereof.

In a further preferred embodiment of the invention, there are provided chimeric  
15 adenoviral vectors comprising nucleotide sequence of a first adenovirus, wherein a portion of a gene thereof encoding a protein that facilitates binding of said vector to a target mammalian cell, or internalization thereof within said cell, is replaced by a portion of the corresponding gene from a second adenovirus belonging to subgroup D, said vectors further comprising a transgene operably linked to a eucaryotic promoter to  
20 allow for expression therefrom in a mammalian cell. In a representative aspect thereof, the replaced encoding sequence codes for a portion of the Ad fiber, hexon or penton base proteins, or combinations thereof. Where a portion of a gene from a second adenovirus is used to construct a chimeric adenoviral vector, such sequence will have a length sufficient to confer a desired serotypic-specific virus-cell interaction to the  
25 vector.

The present invention involves the recognition that adenoviral vectors that are either based substantially upon the genome of Ad serotypes classified in subgroup D, or that contain certain Ad-protein encoding polynucleotide sequences of subgroup D adenovirus, are particularly effective at binding to, and internalizing within, human

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cells, such that therapeutic transgenes included in the adenoviral vector are efficiently expressed. This discovery is particularly surprising given that adenovirus serotypes of subgroup D are not clinically associated with human respiratory disease, and that, for example association with conjunctivitis is more typical. The recognition of this tropism is of particular relevance for the treatment by gene therapy of recognized disease states such as cystic fibrosis or  $\alpha$  1-antitrypsin deficiency. This discovery is particularly surprising given that adenovirus serotypes of subgroup D are not clinically associated with human respiratory disease, and that, for example association with conjunctivitis is more typical. The recognition of this tropism is of particular relevance for the treatment by gene therapy of recognized disease states such as cystic fibrosis or  $\alpha$  1-antitrypsin deficiency.

In a representative aspect of the invention, the adenoviral vectors further comprise nucleotide sequences coding for one or more transgenes and have enhanced tropism for airway epithelial cells. Preferably, the chimeric adenoviral vectors are replication-defective, a feature which contributes to the enhanced safety of adenoviral vectors administered to individuals.

Preferably, the second adenovirus is a member of subgroup D, and the replaced nucleotide sequence encodes a polypeptide selected from the group consisting of Ad fiber, a fragment of Ad fiber, Ad hexon, a fragment of Ad hexon, Ad penton base, and a fragment of Ad penton base. In a preferred embodiment, said second adenovirus is selected from the group consisting of serotypes Ad 9, Ad 15, Ad 17, Ad 19, Ad 20, Ad 22, Ad 26, Ad 27, Ad 28, Ad 30, and Ad 39. In a most preferred embodiment, the second adenovirus is Ad 17. In other preferred embodiments of the chimeric adenoviral vectors, the first adenovirus is selected from the group consisting of Ad 2, Ad 5, and Ad 12.

There is substantial evidence that any reported transforming properties of the E4 region of certain subgroup D serotypes do not extend to Ad serotypes whose use is preferred according to the practice of the present invention (see, for example, R. Javier

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et al., Science, 257, 1992, pp. 1267-1271). It is expected also that, for example, individual ORFs of subgroup D E4 region, such as ORF1, could be deleted.

Additional aspects of the invention include methods to provide biologically active and/or therapeutic proteins to mammalian cells, including, but not limited to, the airway epithelial cells of individuals, in order to provide phenotypic benefit. According to this aspect of the invention, chimeric adenoviral vectors are used in which a nucleotide sequence of a first adenovirus is replaced by the corresponding nucleotide sequence of a second adenovirus. Preferably, the second adenovirus is a member of subgroup D, and the replaced nucleotide sequence encodes a polypeptide encoding all or part of Ad fiber, Ad hexon, or Ad penton base, or combinations thereof.

A still further representative aspect of the invention involves providing a biologically active and/or therapeutic protein in the airway epithelial cells of a patient by administering to said cells an adenoviral vector comprising elements of an Ad 17 genome, and a transgene encoding said protein that is operably linked to a eucaryotic promoter to allow for expression therefrom in a mammalian cell, under conditions whereby the transgene encoding said protein is expressed, and the desired phenotypic benefit is produced in said airway epithelial cells. According to the practice of the invention, it is preferred that an chimeric adenovirus vector utilized to deliver a transgene to the respiratory epithelium (including that of the nasal airway, trachea, and bronchi and alveoli of the lung), or to other tissues of the body, comprise serotypes within subgroup D, as such classification is recognized in the art.

In order to construct the chimeric adenoviral vectors of the invention, reference may be made to the substantial body of literature on how such vectors may be designed, constructed and propagated using techniques from molecular biology and microbiology that are well-known to the skilled artisan. Specific examples of adenoviral vector genomes which can be used as the backbone for a chimeric adenoviral vector of the invention include, for example, Ad2/CFTR-1 and Ad2/CFTR-2 and others described in U. S. Patent No. 5,670,488, issued September 23, 1997

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(incorporated herein by reference). Such vectors may include deletion of the E1 region, partial or complete deletion of the E4 region, and deletions within, for example, the E2 and E3 regions. Within the scope of the invention are, for example, chimeric vectors which contain an Ad 2 backbone with one or more Ad 17 capsid proteins or fragments thereof in the virus. Other adenoviral vector genomic designs which can be used in the chimeric adenoviral vectors of the invention include those derived from allowed U.S. Patent Application Serial No. 08/409,874, filed March 24, 1995, and allowed U.S. Patent Application Serial No. 08/540,077, filed October 6, 1995 (both incorporated herein by reference).

10 To construct the recombinant chimeric adenoviral vectors of the invention which contain a transcription unit, the skilled artisan can use the standard techniques of molecular biology to engineer a transgene or a capsid protein into a backbone vector genome (Berkner, K.L., Curr. Top. Micro. Immunol. 158:39-66, 1992). For example, a plasmid containing a transgene and any operably linked regulatory elements inserted into an adenovirus genomic fragment can be co-transfected with a linearized viral genome derived from an adenoviral vector of interest into a recipient cell under conditions whereby homologous recombination occurs between the genomic fragment and the virus. Preferably, a transgene is engineered into the site of an E1 deletion. As a result, the transgene is inserted into the adenoviral genome at the site in which it was cloned into the plasmid, creating a recombinant adenoviral vector. 20 The chimeric adenoviral vectors can also be constructed using standard ligation techniques, for example, removing a restriction fragment containing a fiber gene from a first adenovirus and ligating into that site a restriction fragment containing a fiber gene from a second adenovirus. A representative example of a chimeric adenoviral vector of the invention is Ad2/ $\beta$ gal-2 fiber 17 (exemplified in Example 6).

25 Construction of the chimeric adenoviral vectors can be based on adenovirus DNA sequence information widely available in the field, e.g., nucleic acid sequence databases such as GenBank.



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Preparation of replication-defective chimeric adenoviral vector stocks can be accomplished using cell lines that complement viral genes deleted from the vector, e.g., 293 or A549 cells containing the deleted adenovirus E1 genomic sequences. The use of HER3 cells (human embryonic retinoblasts transformed by Ad 12), as a  
5 complementing cell line is of note. After amplification of plaques in suitable complementing cell lines, the viruses can be recovered by freeze-thawing and subsequently purified using cesium chloride centrifugation. Alternatively, virus purification can be performed using chromatographic techniques, e.g., as set forth in International Application No. PCT/US96/13872, filed August 30, 1996, incorporated  
10 herein by reference.

Titers of replication-defective chimeric adenoviral vector stocks can be determined by plaque formation in a complementing cell line, e.g., 293 cells. End-point dilution using an antibody to the adenoviral hexon protein may be used to quantitate virus production or infection efficiency of target cells (Armentano et al.,  
15 Hum. Gene Ther. 6:1343-1353, 1995, incorporated herein by reference).

Transgenes which can be delivered and expressed from a chimeric adenoviral vector of the invention include, but are not limited to, those encoding enzymes, blood derivatives, hormones, lymphokines such as the interleukins and interferons, coagulants, growth factors, neurotransmitters, tumor suppressors, apolipoproteins,  
20 antigens, and antibodies, and other biologically active proteins. Specific transgenes which may be encoded by the chimeric adenoviral vectors of the invention include, but are not limited to, cystic fibrosis transmembrane regulator (CFTR), dystrophin, glucocerebrosidase, tumor necrosis factor, p53, p21, herpes simplex thymidine kinase and gancyclovir, retinoblastoma (Rb), and adenosine deaminase (ADA). Transgenes  
25 encoding antisense molecules or ribozymes are also within the scope of the invention. The vectors may contain one or more transgenes under the control of one or more regulatory elements.

In addition to containing the DNA sequences encoding one or more transgenes, the chimeric adenoviral vectors of the invention may contain any



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expression control sequences such as a promoter or enhancer, a polyadenylation element, and any other regulatory elements that may be used to modulate or increase expression, all of which are operably linked in order to allow expression of the transgene. The use of any expression control sequences, or regulatory elements, which facilitate expression of the transgene is within the scope of the invention. Such sequences or elements may be capable of generating tissue-specific expression or be susceptible to induction by exogenous agents or stimuli.

Infection of target cell by the chimeric adenoviral vectors of the invention may also be facilitated by the use of cationic molecules, such as cationic lipids as disclosed in PCT Publication No. WO96/18372, published June 20, 1996, incorporated herein by reference.

Cationic amphiphiles have a chemical structure which encompasses both polar and non-polar domains so that the molecule can simultaneously facilitate entry across a lipid membrane with its non-polar domain while its cationic polar domain attaches to a biologically useful molecule to be transported across the membrane.

Cationic amphiphiles which may be used to form complexes with the chimeric adenoviral vectors of the invention include, but are not limited to, cationic lipids, such as DOTMA (Felgner et al., Proc. Natl. Acad. Sci. USA 84:7413-7417, 1987) (N-[1-(2,3-dioletoxy)propyl]-N,N,N - trimethylammonium chloride); DOGS (dioctadecylamidoglycylspermine) (Behr et al., Proc. Natl. Acad. Sci. USA 86:6982-6986, 1989); DMRIE (1,2-dimyristyloxypropyl-3-dimethyl-hydroxyethyl ammonium bromide) (Felgner et al., J. Biol. Chem. 269:2550-2561, 1994; and DC-chol (3B [N-N', N'-dimethylaminoethane) -carbamoyl] cholesterol) (U.S. Patent No. 5, 283,185 to Epanand et al.). The use of other cationic amphiphiles recognized in the art or which come to be discovered is within the scope of the invention.

In preferred embodiments of the invention, the cationic amphiphiles useful to complex with and facilitate transfer of the vectors of the invention are those lipids which are described in PCT Publication No. WO96/18372, published June 20, 1996, which is incorporated herein by reference. Preferred cationic amphiphiles described

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herein to be used in the delivery of the plasmids and/or viruses are GL-53, GL-67, GL-75, GL-87, GL-89, and GL-120, including protonated, partially protonated, and deprotonated forms thereof. Further embodiments include the use of non-T-shaped amphiphiles as described on pp. 22-23 of the aforementioned PCT application, including protonated, partially protonated and deprotonated forms thereof. Most preferably, the cationic amphiphile which can be used to deliver the vectors of the invention is spermine cholesterol carbamate (GL-67).

In the formulation of compositions comprising the chimeric adenoviral vectors of the invention, one or more cationic amphiphiles may be formulated with neutral co-lipids such as dileoylphosphatidylethanolamine (DOPE) to facilitate delivery of the vectors into a cell. Other co-lipids which may be used in these complexes include, but are not limited to, diphytanoylphosphatidylethanolamine, lyso-phosphatidylethanolamines, other phosphatidylethanolamines, phosphatidylcholines, lyso-phosphatidylcholines and cholesterol. A preferred molar ratio of cationic amphiphile to colipid is 1:1. However, it is within the scope of the invention to vary this ratio, including also over a considerable range. In a preferred embodiment of the invention, the cationic amphiphile GL-67 and the neutral co-lipid DOPE are combined in a 1:2 molar ratio, respectively, before complexing with a chimeric adenoviral vector for delivery to a cell.

In the formulation of complexes containing a cationic amphiphile with a chimeric adenoviral vector, a preferred range of  $10^7$  -  $10^{10}$  infectious units of virus may be combined with a range of  $10^4$  -  $10^6$  cationic amphiphile molecules/viral particle.

The infection efficiency of the chimeric adenoviral vectors of the invention may be assayed by standard techniques to determine the infection of target cells. Such methods include, but are not limited to, plaque formation, end-point dilution using, for example, an antibody to the adenoviral hexon protein, and cell binding assays using radiolabelled virus. Improved infection efficiency may be characterized as an increase in infection of at least an order of magnitude with reference to a control virus. Where

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a chimeric adenoviral vector encodes a marker or other transgene, relevant molecular assays to determine expression include the measurement of transgene mRNA, by, for example, Northern blot, S1 analysis or reverse transcription-polymerase chain reaction (RT-PCR). The presence of a protein encoded by a transgene may be detected by  
5 Western blot, immunoprecipitation, immunocytochemistry, or other techniques known to those skilled in the art. Marker-specific assays can also be used, such as X-gal staining of cells infected with a chimeric adenoviral vector encoding  $\beta$ -galactosidase.

In order to determine transgene expression and infection efficiency in vivo using the constructs and compositions of the invention, animal models may be  
10 particularly relevant in order to assess transgene persistence against a background of potential host immune response. Such a model may be chosen with reference to such parameters as ease of delivery, identity of transgene, relevant molecular assays, and assessment of clinical status. Where the transgene encodes a protein whose lack is associated with a particular disease state, an animal model which is representative of  
15 the disease state may optimally be used in order to assess a specific phenotypic result and clinical improvement. However, it is also possible that particular chimeric adenoviral vectors of the invention display enhanced infection efficiency only in human model systems, e.g., using primary cell cultures, tissue explants, or permanent cell lines. In such circumstances where there is no animal model system available in  
20 which to model the infection efficiency of a chimeric adenoviral vector with respect to human cells, reference to art-recognized human cell culture models will be most relevant and definitive.

Relevant animals in which the chimeric adenoviral vectors may be assayed include, but are not limited to, mice, rats, monkeys, and rabbits. Suitable mouse  
25 strains in which the vectors may be tested include, but are not limited to, C3H, C57Bl/6 (wild-type and nude) and Balb/c (available from Taconic Farms, Germantown, New York).

Where it is desirable to assess the host immune response to vector administration, testing in immune-competent and immune-deficient animals may be

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compared in order to define specific adverse responses generated by the immune system. The use of immune-deficient animals, e.g., nude mice, may be used to characterize vector performance and persistence of transgene expression, independent of an acquired host response.

5           In a particular embodiment where the transgene is the gene encoding cystic fibrosis transmembrane regulator protein (CFTR) which is administered to the respiratory epithelium of test animals, expression of CFTR may be assayed in the lungs of relevant animal models, for example, C57Bl/6 or Balb/c mice, cotton rats, or Rhesus monkeys. Molecular markers which may be used to determine expression  
10       include the measurement of CFTR mRNA, by, for example, Northern blot, S1 analysis or RT-PCR. The presence of the CFTR protein may be detected by Western blot, immunoprecipitation, immunocytochemistry, or other techniques known to those skilled in the art. Such assays may also be used in tissue culture where cells deficient in a functional CFTR protein and into which the chimeric adenoviral vectors have  
15       been introduced may be assessed to determine the presence of functional chloride ion channels - indicative of the presence of a functional CFTR molecule.

          The chimeric adenoviral vectors of the invention have a number of in vivo and in vitro utilities. The vectors can be used to transfer a normal copy of a transgene encoding a biologically active protein to target cells in order to remedy a deficient or  
20       dysfunctional protein. The vectors can be used to transfer marked transgenes (e.g., containing nucleotide alterations) which allow for distinguishing expression levels of a transduced gene from the levels of an endogenous gene. The chimeric adenoviral vectors can also be used to define the mechanism of specific viral protein-cellular protein interactions that are mediated by specific virus surface protein sequences. The  
25       vectors can also be used to optimize infection efficiency of specific target cells by adenoviral vectors, for example, using a chimeric adenoviral vector containing Ad 17 fiber protein to infect human nasal polyp cells. Where it is desirable to use an adenoviral vector for gene transfer to cancer cells in an individual, a chimeric adenoviral vector can be chosen which selectively infects the specific type of target

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cancer cell and avoids promiscuous infection. Where primary cells are isolated from a tumor in an individual requiring gene transfer, the cells may be tested against a panel of chimeric adenoviral vectors to select a vector with optimal infection efficiency for gene delivery. The vectors can further be used to transfer tumor antigens to dendritic  
5 cells which can then be delivered to an individual to elicit an anti-tumor immune response. Chimeric adenoviral vectors can also be used to evade undesirable immune responses to particular adenovirus serotypes which compromise the gene transfer capability of adenoviral vectors.

The present invention is further directed to compositions containing the  
10 chimeric adenoviral vectors of the invention which can be administered in an amount effective to deliver one or more desired transgenes to the cells of an individual in need of such molecules and cause expression of a transgene encoding a biologically active protein to achieve a specific phenotypic result. The cationic amphiphile-plasmid complexes or cationic amphiphile-virus complexes may be formulated into  
15 compositions for administration to an individual in need of the delivery of the transgenes.

The compositions can include physiologically acceptable carriers, including any relevant solvents. As used herein, "physiologically acceptable carrier" includes any and all solvents, dispersion media, coatings, antibacterial and antifungal agents,  
20 isotonic and absorption delaying agents, and the like. Except insofar as any conventional media or agent is incompatible with the active ingredient, its use in the compositions is contemplated.

Routes of administration for the compositions containing the chimeric adenoviral vectors of the invention include conventional and physiologically  
25 acceptable routes such as direct delivery to a target organ or tissue, intranasal, intravenous, intramuscular, subcutaneous, intradermal, oral and other parenteral routes of administration.

The invention is further directed to methods for using the compositions of the invention in vivo or ex vivo applications in which it is desirable to deliver one or more

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transgenes into cells such that the transgene produces a biologically active protein for a normal biological or phenotypic effect. In vivo applications involve the direct administration of one or more chimeric adenoviral vectors formulated into a composition to the cells of an individual. Ex vivo applications involve the transfer of  
5 a composition containing the chimeric adenoviral vectors directly to autologous cells which are maintained in vitro, followed by readministration of the transduced cells to a recipient.

Dosage of the chimeric adenoviral vector to be administered to an individual for expression of a transgene encoding a biologically active protein and to achieve a  
10 specific phenotypic result is determined with reference to various parameters, including the condition to be treated, the age, weight and clinical status of the individual, and the particular molecular defect requiring the provision of a biologically active protein. The dosage is preferably chosen so that administration causes a specific phenotypic result, as measured by molecular assays or clinical markers. For  
15 example, determination of the infection efficiency of a chimeric adenoviral vector containing the CFTR transgene which is administered to an individual can be performed by molecular assays including the measurement of CFTR mRNA, by, for example, Northern blot, S1 or RT-PCR analysis or the measurement of the CFTR protein as detected by Western blot, immunoprecipitation, immunocytochemistry, or  
20 other techniques known to those skilled in the art. Relevant clinical studies which could be used to assess phenotypic results from delivery of the CFTR transgene include PFT assessment of lung function and radiological evaluation of the lung. Demonstration of the delivery of a transgene encoding CFTR can also be  
25 demonstrated by detecting the presence of a functional chloride channel in cells of an individual with cystic fibrosis to whom the vector containing the transgene has been administered (Zabner et al., J. Clin. Invest. 97:1504-1511, 1996). Transgene expression in other disease states can be assayed analogously, using the specific clinical parameters most relevant to the condition.



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Dosages of a chimeric adenoviral vector which are effective to provide expression of a transgene encoding a biologically active protein and achieve a specific phenotypic result range from approximately  $10^8$  infectious units (I.U.) to  $10^{11}$  I.U. for humans.

5 It is especially advantageous to formulate parenteral compositions in dosage unit form for ease of administration and uniformity of dosage. Dosage unit form as used herein refers to physically discrete units suited as unitary dosages for the subjects to be treated, each unit containing a predetermined quantity of active ingredient calculated to produce the specific phenotypic effect in association with the required  
10 physiologically acceptable carrier. The specification for the novel dosage unit forms of the invention are dictated by and directly depend on the unique characteristics of the chimeric adenoviral vector and the limitations inherent in the art of compounding. The principal active ingredient (the chimeric adenoviral vector) is compounded for convenient and effective administration in effective amounts with the physiologically  
15 acceptable carrier in dosage unit form as discussed above.

Maximum benefit and achievement of a specific phenotypic result from administration of the chimeric adenoviral vectors of the invention may require repeated administration. Such repeated administration may involve the use of the same chimeric adenoviral vector, or, alternatively, may involve the use of different  
20 chimeric adenoviral vectors which are rotated in order to alter viral antigen expression and decrease host immune response.

The practice of the invention employs, unless otherwise indicated, conventional techniques of protein chemistry, molecular virology, microbiology, recombinant DNA technology, and pharmacology, which are within the skill of the  
25 art. Such techniques are explained fully in the literature. See, e.g., Current Protocols in Molecular Biology, Ausubel et al., eds., John Wiley & Sons, Inc., New York, 1995, and Remington's Pharmaceutical Sciences, 17th ed., Mack Publishing Co., Easton, PA, 1985.



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The invention is further illustrated by the following specific examples which are not intended in any way to limit the scope of the invention.

### Examples

5

#### Example 1 Infection of NHBE cells by adenovirus serotypes of subgroup D

Normal human bronchial epithelial ("NHBE") cells were obtained from Clonetics (San Diego, CA), and plated on Costar (Cambridge, MA) Transwell-Clear polyester membranes that were pre-coated with human placental collagen. The wells  
10 were placed in a cluster plate and cells were fed every day for one week by changing the medium in both the well and the plate. After one week the media was removed from the wells to create an air-liquid interface, and the cells were then fed only by changing the medium in the cluster plate, every other day for one week. Cells were infected at an moi of 1 by adding virus (see below) to the transwell, followed by an  
15 incubation time of 1.5-2 hours. At the end of the incubation period, the medium was removed and the cells were gently rinsed with fresh medium. Thirty-six hours post-infection the cells were fixed with 1:1 acetone:methanol, permeablized with a solution of 0.05% Tween 20 in PBS, and stained with FITC labeled anti-hexon antibody (Chemicon, Temecula, CA) to visualize cells that had been productively infected (i.e.  
20 to visualize virus replication). Cells were also subjected to the DAPI staining procedure in order to visualize the total number of nuclei. The results could be readily determined upon simple inspection.

Wild type Ad serotypes within subgroup D that were tested included 9, 15, 17, 19, 20, 22, 26, 27, 28, 30, and 39 (all from the American Type Culture Collection,  
25 Rockville, MD). An Ad 2 (obtained as DNA from BRL, Gaithersburg, MD, and used to transfect 293 cells in order to generate virus stock) was used as a control. Infection observed with all of the subgroup D serotypes was superior to that observed with Ad 2, with the best results being achieved with Ad 9, Ad 17, Ad 20, Ad 22, and Ad 30.

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Additionally, it was determined that each of the above-mentioned serotypes of subgroup D was more effective in the NHBE cell assay under similar circumstances than any other serotype tested than belongs to a subgroup other than D. In this regard, the following serotypes were also tested: 31(subgroup A); 3(subgroup B); 7(subgroup B); 7a(subgroup B); 14(subgroup B); 4(subgroup E); and 41(subgroup F). In a further experiment, serotype 35 (subgroup A) may have performed as well as the least effective members of subgroup D that were tested.

#### Example 2 Infection of clinical isolate bronchial epithelial cells

Following generally the procedures of Example 1, human bronchial epithelial cells recovered from healthy human volunteers were infected with either Ad 2 (as above, Ad 2 DNA was obtained from BRL, and this DNA was used to transfect 293 cells to generate virus) (Figure 1), or Ad 17 (from ATCC) (Figure 2), all at an moi of 50. Cells were left in contact with virus for 30 minutes, 3 hours, or 12 hours.

The increased tropism of Ad 17 for human bronchial epithelial cells, compared with Ad 2, is readily apparent upon inspection of Figures 1 and 2. In the Figures, the right hand columns (panels D, E, and F, stained in blue) show total numbers of cells present (from DAPI staining as above), whereas the left hand columns (panels A, B, and C, stained in green) quantify adenovirus hexon protein present in the infected cells (from FITC-labeled anti-hexon antibody, as above). Panels A and D result from 30 minute incubation times, panels B and E result from 3 hour incubation times, and panels C and F result from 12 hour incubation times. As measured by the technique employed, infection of airway epithelia by Ad 17 is at least 50 fold greater than by Ad 2 for the thirty minute incubation time.

#### Example 3 Binding of Ad 2 and Ad 17 to human nasal polyp cell isolates

293 cells, a complementing cell line developed by Graham et al. (see Gen. Virol. , 36, 1977, pp. 59-72), were infected with either wild type Ad 2 or wild type Ad 17. Five hours post-infection the media was removed and replaced with methionine

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free media containing  $S^{35}$  metabolic label (Amersham). After an additional six hours, fresh media was added and the labeling was allowed to proceed for a total of 18 hours, after which the  $S^{35}$  media was removed and replaced with fresh media. Thirty hours post-infection the cells were harvested and lysed and the labeled Ad 2 or Ad 17  
5 viruses were purified by CsCl gradient centrifugation. The recovered viruses were then used in an assay to determine their relative binding efficiency on human nasal polyp cells.

In order to perform the assay, ciliated human airway epithelial cells were recovered from nasal polyps of healthy volunteers. The results from two such isolates,  
10 NP-14 and NP-15, are reported here (see Figure 3). Radiolabeled virus was then incubated with the isolated cells in wells for specified times ( 5 or 30 minutes, see Figure 3). The cells were then rinsed and measured for radioactivity. Binding as reported in Figure 3 indicates the percent of input radioactivity that is cell associated. It was determined that for both cell isolate populations, using either 5 or 30 minute  
15 incubations, cell associated radioactivity was 10-fold enhanced if Ad 17 rather than Ad 2 was used.

#### Example 4 Fiber competition

20 A549 cells (a human lung carcinoma line, obtained from the American Type Culture Collection as ATCC CCL-185) were plated at  $3 \times 10^4$  cells per well in 96-well dishes. Since the number of receptor sites for adenovirus fiber on the cell surface has been estimated to be approximately  $10^5$  receptors per cell, the receptors in the plated cells were saturated, in this example, with  $0.1 \mu\text{g}$  of purified full length Ad 2 fiber  
25 protein (obtained from Paul Freimuth, Brookhaven National Laboratory, Upton, NY), which corresponds to approximately 100 molecules of fiber per receptor. Cells were incubated with Ad 2 fiber in PBS for two hours at  $37^\circ\text{C}$ .

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The cells were subsequently infected at an moi of 1 (using either Ad 2 provided as above, or wild type Ad 17) for one hour, after which the cells were rinsed, and fresh medium was added. Control cultures were incubated with PBS with no added protein for two hours and then subsequently infected as described above. Forty  
5 hours post-infection the cells were fixed with 1:1 acetone:methanol, permeabilized with 0.05% Tween 20 in PBS and stained with FITC labeled anti- Ad 2 hexon antibody, as described in Example 1. As determined by this assay, the number of cells infected (stained) with Ad 2 was reduced by approximately 90% in cultures that were pre-incubated with Ad 2 fiber as compared to control cultures. However, no effect on  
10 Ad 17 infection was observed by the pre-incubation of A549 cells with full length Ad 2 fiber.

Example 5 Use of Ad 2 fiber knob in a binding competition  
experiment with Ad 2

15

Further competition experiments were performed with Ad 2 and Ad 17 fiber knobs that had been expressed and purified from *E. coli*. DNA sequences encoding both protein fragments were designed so that the fiber knobs expressed therefrom would contain histidine tags in order to permit nickel- column purification. The yield  
20 of soluble fiber knob trimer, purified by the Ni-NTA method (Qiagen, Chatsworth, CA), was ~25µg/50ml culture. A significant portion of the total knob protein expressed appeared to remain in a monomeric (and insoluble) form. The soluble trimeric material obtained was used for a preliminary competition experiment. Wild type Ad 2 and Ad 17 were used to infect A549 cells, or cells that had been pre-  
25 incubated with excess (about 100 molecules of trimer per receptor) Ad 2 fiber knob or Ad 17 fiber knob. The results indicated that Ad 2 fiber knob, but not Ad 17 knob, could block Ad 2 infection. Additionally, Ad 17 infection was not blocked by *E. coli*-expressed fiber knobs of either serotype, suggesting that the mechanism of Ad 2 and Ad 17 infections is different.

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Example 6 Construction of the chimeric vector Ad2/ $\beta$ gal-2/fiber Ad 17

The vector Ad2/ $\beta$ gal-2 was constructed as follows. A CMV $\beta$ gal expression  
5 cassette was constructed in a pBR322-based plasmid that contained Ad 2 nucleotides  
1-10,680 from which nucleotides 357-3328 were deleted. The deleted sequences were  
replaced with (reading from 5' to 3'): a cytomegalovirus immediate early promoter  
(obtained from pRC/CMV, Invitrogen), lacZ gene encoding  $\beta$ -galactosidase with a  
nuclear localization signal, and an SV40 polyadenylation signal (nucleotides 2533-  
10 2729). The resulting plasmid was used to generate Ad2/ $\beta$ gal-2 by recombination with  
Ad2E4ORF6 (D. Armentano et al., Human Gene Therapy , 6, 1995, pp 1343 -1353).

A chimeric Ad2/ $\beta$ gal-2/fiber Ad 17 viral vector (Figure 4) was then constructed  
as follows. pAdORF6 (D. Armentano et al., Human Gene Therapy , 6, 1995, pp 1343  
-1353 was cut with Nde and BamHI to remove Ad 2 fiber coding and polyadenylation  
15 signal sequences (nucleotides 20624-32815). An NdeI-BamHI fragment containing  
Ad 17 fiber coding sequence (nucleotides 30984-32095) was generated by PCR and  
ligated along with an SV40 polyadenylation signal into NdeI-BamHI cut pAdORF6 to  
generate pAdORF6fiber17. This plasmid was cut with PacI and then ligated to PacI-  
cut Ad2/ $\beta$ gal-2 DNA to generate Ad2/ $\beta$ gal-2fiber 17. Any desired transgene may be  
20 substituted in this construct for the reporter gene.

A similar construct can be prepared using a DNA sequence that encodes Ad 17  
penton base instead of Ad 17 fiber. Alternatively, only a subregion of the penton base  
of Ad 2 need be subject to replacement, such as by inserting into the vector a  
nucleotide encoding sequence corresponding to any amino acid subsequence of Ad 17  
25 penton base amino acids 283-348 ( see the marked sequence in Figure 5A) in  
replacement for any subsequence of Ad 2 penton base amino acids 290-403.  
Preferrably, the replaced sequence of Ad 2 and the inserted sequence of Ad 17  
includes the RGD domain of each. Use of nucleotide sequence corresponding to  
penton base amino acid sequence for other subgroup D serotypes is also within the

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practice of the invention. It is also within the scope of the invention to replace a subregion of the fiber protein in the Ad 2 vector with a subregion from another adenovirus serotype, for example, Ad 17.

5 Example 7 Ad2/ $\beta$ gal-2f17 shows increased infection efficiency on human airway explants

Both human and monkey trachea explants, about 1 cm<sup>2</sup>, were placed on top of an agar support. Each explant was infected at an moi of 200 of either Ad2/ $\beta$ gal-2 or Ad2/ $\beta$ gal-2f17 assuming a cell density of 1 x 10<sup>6</sup> per cm<sup>2</sup> of explant. Explants were  
10 exposed to virus for three hours and were then rinsed with NHBE media. Two days post-infection explants were stained with X-gal and infection efficiency was assessed. On the monkey explants Ad2/ $\beta$ gal-2 gave rise to a higher infection efficiency than Ad2/ $\beta$ gal-2f17. Patches of stained cells were detected in explants exposed to Ad2/ $\beta$ gal-2 but very few cells stained in explants exposed to Ad2/ $\beta$ gal-2f17. A  
15 different result was obtained on human trachea explants. On these explants Ad2/ $\beta$ gal-2f17 infection gave rise to a much higher infection efficiency than Ad2/ $\beta$ gal-2 infection. Approximately 5-10% of the cells in explants exposed to Ad2/ $\beta$ gal-2f17 stained with X-gal whereas very few cells were stained in explants exposed to Ad2/ $\beta$ gal-2. No background staining was observed in either monkey or human  
20 explants that were not exposed to virus.

The results indicate that the exchange of Ad 2 fiber for Ad 17 fiber in Ad2/ $\beta$ gal-2f17 was sufficient to significantly increase infection efficiency of human tracheal airway cells by an adenovirus type 2 based vector.

25 Example 8 Adenovirus subgroup screening on human cancer cell lines

Identification of adenovirus subgroup that best infects a particular tumor type may be useful in designing vectors to optimally target cancer cells in vivo. In order to determine the adenovirus subgroup that best infects a particular type of cancer cell, cancer cells were seeded into a 96 well plate and infected with and moi of 5. Infection

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efficiency was determined by staining of infected cells using an anti-hexon antibody. The adenovirus subgroups were represented by the following serotypes: A: Ad 31; B: Ad 3; C: Ad 2; D: Ad 17; E: Ad 4; and F: Ad 41.

Subgroup D (Ad 17) has a significantly higher infection rate of the colon  
5 cancer cell line CaCo-2 than other cell types, with an infection rate of 70%, while Ad 2 only infected 20% of the cells (Figure 9).

Subgroup D (Ad 17) was effective in infecting ovarian cancer cell line SK-OV3. Infection was measured at 90% (Figure 10).

#### 10 Sequence Listing

Included herewith on the following pages are informal copies of SEQ ID NO: 1 through SEQ ID NO: 3.



1 CATCATCAAT AATATACCCC ACAAAGTAAA CAAAAGTTAA TATGCAAATG AGGTTTTTAAA  
 61 TTTAGGGCGG GGCTACTGCT GATTGGCCGA GAAACGTTGA TGCAAATGAC GTCACGACGC  
 121 ACGGCTAACG GTCGCCGCGG AGGCGTGGCC TAGCCCGGAA GCAAGTCGCG GGGCTGATGA  
 181 CGTATAAAAA AGCGGACTTT AAACCCGGAA ACGGCCGATT TTCCCGCGGC CACGCCCGGA  
 241 TATGAGGTAA TTCTGGGCGG ATGCAAGTGA AATTAGGTCA TTTTGGCGCG AAAACTGAAT  
 301 GAGGAAGTGA AAAGTGAAAA ATACCGGTCC CGCCCAGGGC GGAATATTTA CCGAGGGCCG  
 361 AGAGACTTTG ACCGATTACG TGTGGGTTTC GATTGCGGTG TTTTTCGCG AATTTCGCG  
 421 TCCGTGTCAA AGTCCGGTGT TTATGTCACA GATCAGCTGA TCCACAGGGT ATTTAAACCA  
 481 GTCGAGCCCG TCAAGAGGCC ACTCTTGAGT GCCAGCGAGT AGAGATTTCT CTGAGCTCCG  
 541 CTCCCAGAGT GTGAGAAAAA TGAGACACCT GCGCCTCCTG CCTGGAAGTG TGCCCTTGA  
 601 CATGGCCGCA TTATTGCTGG ATGACTTTGT GAGTACAGTA TTGGAGGATG AACTGCAACC  
 661 AACTCCGTTT GAGCTGGGAC CCACACTTCA GGACCTCTAT GATTGGAGG TAGATGCCCA  
 721 GGAGGACGAC CCGAACGAAG ATGCTGTGAA TTTAATATTT CCAGAATCTC TGATTCTTCA  
 781 GGCTGACATA GCCAGCGAAG CTCTACCTAC TCCACTTCAT ACTCCAATC TGTCACCCAT  
 841 ACCTGAATTG GAAGAGGAGG ACGAGTTAGA CCTCCGGTGT TATGAGGAAG GTTTTCTCC  
 901 CAGCGATTCA GAGGACGAAC AGGGTGAGCA GAGCATGGCT CTAATCTCAG ACTATGCTTG  
 961 TGTGGTTGTG GAAGAGCATT TTGTGTTGGA CAATCCTGAG GTGCCCGGGC AAGGCTGTAA  
 1021 ATCCTGCCAG TACCACCGGG ATAAGACCGG AGACACGAAC GCCTCCTGTG CTCTGTGTTA  
 1081 CATGAAAAAG AACTTCAGCT TTATTTACAG TAAGTGGAGT GAATGTGAGA GAGGCTGAGT  
 1141 GCTTAAGACA TAACTGGGTG ATGCTTCAAC AGCTGTGCTA AGTGTGGTTT ATTTTGTTC  
 1201 TAGGTCCGGT GTCAGAGGAT GGTCAATCACC CTCAGAAGAA GACCACCCGT GTCCCCCTGA  
 1261 TCTGTCAGGC GAAACGCCCC TGCAAGTGCA CAGACCCACC CCAGTCAGAC CCAGTGGCGA  
 1321 GAGGCGAGCA GCTGTTGAAA AAATTGAGGA CTTGTTACAT GACATGGGTG GGGATGAACC  
 1381 TTTGGACCTG AGCTTGAAAC GTCCCAGGAA ACTAGGCGCA GCTGCGCTTA GTCATGTGTA  
 1441 AATAAAGTTG TACAATAAAA ATTATATGTG ACGCATGCAA GGTGTGGTTT ATGACTCATG  
 1501 GGCGGGGCTT AGTTCTATAT AAGTGGCAAC ACCTGGGCAC TGGAGCACAG ACCTTCAGGG  
 1561 AGTTCCTGAT GGATGTGTGG ACTATCCTTG CAGACTTTAG CAAGACACGC CGGCTGTAG  
 1621 AGGATAGTTC AGACGGGTGC TCCGGGTCTT GGAGACACTG GTTTGGAACCT CCTCTATCTC  
 1681 GCCTGGTGTG CACAGTTAAA AAGGATTATA ACGAGGAATT TGAAAATCTT TTTGCTGATT  
 1741 GCTCTGGCCT GCTAGATTCT CTGAATCTCG GCCACCAGTC CCTTTTCCAG GAAAGGGTAC  
 1801 TCCACAGCCT TGATTTTTCC AGCCAGGGC GCACTACAGC CGGGGTGCT TTTGTGGTTT  
 1861 TTCTGGTTGA CAAATGGAGC CAGAACACCC AACTGAGCAG GGGCTACATT CTGGACTTCG  
 1921 CAGCCATGCA CCTGTGGAGG GCATGGGTCA GGCAGCGGGG ACAGAGAATC TTGAACACT  
 1981 GGCTTCTACA GCCAGCAGCT CCGGGTCTTC TTCGTCTACA CAGACAAACA TCCATGTTGG  
 2041 AGGAAGAAAT GAGGCAGGCC ATGGACGAGA ACCCGAGGAG CGGTCTGGAC CCTCCGTCGG  
 2101 AAGAGGAGTT GGATTGAATC AGGTATCCAG CCTGTACCCA GAGCTTAGCA AGGTGCTGAC  
 2161 ATCCATGGCC AGGGGAGTGA AGAGGGAGAG GAGCGATGGG GGCAATACCG GGATGATGAC  
 2221 CGAGCTGACG GCCAGTCTGA TGAATCGCAA GCGCCCAGAG CGCCTTACCT GGTACGAGCT  
 2281 ACAGCAGGAG TGCAGGGATG AGTTGGGCCT GATGCAGGAT AAATATGGCC TGGAGCAGAT  
 2341 AAAAACCCAT TGGTTGAACC CAGATGAGGA TTGGGAGGAG GCTATTAAGA AGTATGCCAA  
 2401 GATAGCCCTG CGCCCAGATT GCAAGTACAT AGTGACCAAG ACCGTGAATA TCAGACATGC  
 2461 TGCTACATCT CGGGGAACGG GGCAGAGGTG GTCATTGATA CCCTGGACAA GGCCGCCTTT  
 2521 AGGTGTTGCA TGATGGGAAT GAGAGCCGGA GTGATGAATA TGAATTCCAT GATCTTTATG  
 2581 AACATGAAGT TCAATGGAGA GAAGTTTAAAT GGGGTGCTGT TCATGGCCAA CAGCCACATG  
 2641 ACCCTGCATG GCTGCGACTT TTTCGGCTTT AACAATATGT GCGCAGAGGT CTGGGGCGCT  
 2701 TCCAAGATCA GGGGATGTAA GTTTTATGGC TGCTGGATGG GCGTGGTCGG AAGACCCAAG  
 2761 AGCGAGATGT CTGTGAAGCA GTGTGTGTTT GAGAAATGCT ACCTGGGAGT CTCTACCGAG  
 2821 GGCAATGCTA GAGTGAGGCA CTGCTCTTCC CTGGAGACGG GCTGCTTCTG CCTGGTGAAG  
 2881 GGCACAGCCT CTCTGAAGCA TAATATGGTG AAGGGCTGCA CGGATGAGCG CATGTACAAC  
 2941 ATGCTGACTG CGACTCGGGG GTCTGTCATA TCCTGAAGAA CATCCATGTG ACCTCCACC  
 3001 CCAGAAAGAA GTGGCCAGTG TTTGAGAATA ACATGCTGAT CAAGTGCCAC ATGCACCTGG  
 3061 GCGCCAGAAG GGGCACCTTC CAGCCGTACC AGTGCAACTT TAGCCAGACC AAGCTGCTGT  
 3121 TGGAGAACGA TGCCTTCTCC AGGGTGAACC TGAACGGCAT CTTTGACATG GATGTCTCGG  
 3181 TGTACAAGAT CCTGAGATAC GATGAGACCA AGTCCAGGGT GCGCGCTTGC GAGTGCGGGG  
 3241 GCAGACACAC CAGGATGCAG CCAGTGGCCC TGGATGTGAC CGAGGAGCTG AGACCAGACC  
 3301 ACCTGGTGAT GGCCTGTACC GGGACCGAGT TCAGCTCCAG TGGGGAGGAC ACAGATTAGA  
 3361 GGTAGGTTTG AGTAGTGGGC GTGGCTAAGG TGACTATAAA GGCGGGTGTC TTACGAGGGT

3421 CTTTTTGCTT TTCTGCAGAC ATCATGAACG GGACCGGCGG GGCCTTCGAA GGGGGGCTTT  
3481 TTAGCCCTTA TTTGACAACC CGCCTGCCAG GATGGGCGCG AGTTCGTCAG AATGTGATGG  
3541 GATCGACGGT GGACGGGCGC CCAGTGCTTC CAGCAAATTC CTCGACCATG ACCTACGCGA  
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31741 AGAAGTGACA ATTTAACTGT ATCTGAGGCA TATAAAAATG CAGTTGAATT TATGCCTAAT  
31801 TTGGTAGCCT ACCCAAACC TACCACTGGC TCTAAAAAT ATGCAAGGGA TATAGTCTAT  
31861 GGAACATAT ATCTTGAGG TTTGGCATAT CAGCCAGTTG TAATTAAGGT TACTTTTAAT  
31921 GAAGAAGCAG ATAGTGCTTA CTCTATAACA TTTGAATTTG TATGGAATAA AGAATATGCC  
31981 AGGGTTGAAT TTGAAACCAC TTCCTTTACC TTCTCCTATA TTGCCAACA ATAAAAGACC  
32041 AATAAACGTG TTTTTTATTT CAAATTTTAT GTATCTTTAT TGATTTTAC ACCAGCGCGA  
32101 GTAGTCAATC TCCCACCACC AGCCCATTTT ACAGTGACA CGGTTCTCTC AGCACGGTGG  
32161 CCTTAAATAA GGAAATGTTT TGATTATTGC GGGAACTGGA CTTGGGGTCT ATAATCCACA  
32221 CAGTTTCCTG ACGAGCCAAA CGGGGATCGG TGATTGAAAT GAAGCCGTCC TCTGAAAAGT  
32281 CATCCAAGCG GGCCTCACAG TCCAGGTCAC AGTCTGGTGG AACGAGAAGA ACGCACAGAT  
32341 TCATACTCGG AAAACAGGAT GGGTCTGTGC CTCTCCATCA GCGCCCTCAG CAGTCTCTGC  
32401 CGCCGGGGCT CGGTGCGGCT GCTGCAAATG GGATCGGGAT CACAAGTCTC TCTAACTATG  
32461 ATCCAACAG CCTTCAGCAT CAGTCTCCTG GTGCGTCGAG CACAGCACCG CATCCTGATC  
32521 TCTGCCATGT TCTCACAGTA AGTGCAGCAC ATAATCACCA TGTTATTTCAG CAGCCCATAA  
32581 TTCAGGGTGC TCCAGCCAAA GCTCATGTTG GGGATGATGG AACCACGTG ACCATCGTAC  
32641 CAGATGCGGC AGTATATCAG GTGCCTGCCC CTCATGAACA CACTGCCCAT ATACATGATC  
32701 TCTTTGGGCA TGTTTCTGTT TACAATCTGG CGGTACCAGG GGAAGCGCTG GTTGAACATG  
32761 CACCCGTAAA TGACTCTCCT GAACCACACG GCCAGCAGGG TGCCCTCCCG CCGACACTGC  
32821 AGGGAGCCAG GGGATGAACA GTGGCAATGC AGGATCCAGC GCTCGTACCC GCTCACCATC  
32881 TGAGCTCTTA CCAAGTCCAG GGTAGCGGG CACAGGCACA CTGACATACA TCTTTTAAA  
32941 ATTTTATTTT CCTCTGTGGT GAGGATCATA TCCCAGGGGA CTGGAACTC TTGGAGCAGG  
33001 GTAAAGCCAG CAGCACATGG TAATCCACGG ACAGAACTTA CATTATGATA ATCTGCATGA  
33061 TCACAATCGG GCAACAGGGG ATGTTGATCA GTCAGTGAAG CCCTGGTTTC ATCATCAGAT  
33121 CGTGGTAAAC GGGCCCTGCG ATATGGATGA TGGCGGAGCG AGCTGGATTG AATCTCGGTT  
33181 TGCATTGTAG TGGATTCTCT TGCGTACCTT GTCGTAATTC TGCCAGCAGA AATGGGCCCT  
33241 TGAACAGCAT ATACCCCTCC TGCGGCCGTC CTTTCGCTGC TGCCGCTCAG TCATCCAAC  
33301 GAAGTACATC CATTCTCGAA GATTCTGGAG AAGTTCCTCT GCATCTGATG AAATAAAAAA  
33361 CCCGTCCATG CGAATTCCCC TCATCACATC AGCCAGGACT CTGTAGGCCA TCCCCATCCA  
33421 GTTAATGCTG CCTTGTCTAT CATTACAGAG GGGCGGTGGC AGGATTGGAA GAACCATTTT  
33481 TATTCCAAAC GGTCTCGAAG GACGATAAAG TGCAAGTCAC GCAGGTGACA GCGTTCCCTT  
33541 CCGCTGTGCT GGTGGAAACA GACAGCCAGG TCAAAACCCA CTCTATTTTC AAGGTGCTCG  
33601 ACCGTGGCTT CGAGCAGTGG CTCTACGCGT ACATCCAGCA TAAGAATCAC ATTAAAGGCT  
33661 GGCCCTCCAT CGATTTCATC AATCATCAGG TTACATTCCT GCACCATCCC CAGGTAATTC  
33721 TCATTTTTC AGCCTTGGAT TATCTCTACA AATTGTTGGT GTAAATCCAC TCCGCACATG  
33781 TTGAAAAGCT CCCACAGTGC CCCCTCCACT TTCATAATCA GGCAGACCTT CATAATAGAA  
33841 ACAGATCCTG CTGCTCCACC ACCTGCAGCG TGTTCAAAC AACAAGATTC AATAAGGTTT  
33901 TGCCCTCCGC CCTGAGCTCG CGCCTCAATG TCAGCTGCAA AAAGTCACTT AAGTCCTGGG  
33961 CCACTACAGC TGACAATTCA GAGCCAGGGC TAAGCGTGGG ACTGGCAAGC GTGAGGGAAA  
34021 ACTTTAATGC TCCAAAGCTA GCACCCAAA ACTGCATGCT GGAATAAGCT CTCTTTGTGT  
34081 CTCCGGTGAT GCCTTCCAAA ATGTGAGTGA TAAAGCGTGG TAGTTTTTTC TTTAATCATT  
34141 TGCGTAATAG AAAAGTCCTG TAAATAAGTC ACTAGGACCC CAGGGACCAC AATGTGGTAG

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34201 CTTACACCGC GTCGCTGAAA GCATGGT TAGAGATGAG AGTCTGAAAA ACAGAAAGCA  
34261 TCGCGCTAAAC TAAGGTGGCT ATTTTCACTG AAGGAAAAAT CACTCTTTCC AGCAGCAGGG  
34321 TACCCACTGG GTGGCCCTTG CGGACATACA AAAATCGGTC CGTGTGATTA AAAAGCAGCA  
34381 CAGTAAGTTC CTGTCTTCTT CCGGCAAAAA TCACATCGGA CTGGGT TAGT ATGTCCCTGG  
34441 CATGGTAGTC ATTCAAGGCC ATAAATCTGC CCTGATATCC AGTAGGAACC AGCACACTCA  
34501 CTTTTAGGTG AAGCAATACC ACCCCATGCG GAGGAATGTG GAAAGATTCA GGGCAAAAAA  
34561 AATTATATCT ATTGCTAGCC CTTCTGGAC GGGAGCAATC CTCCAGGACT ATCTATGAAA  
34621 GCATACAGAG ATTCAGCCAT AGCTCAGCCC GCTTACCAGT AGACAAAGAG CACAGCAGTA  
34681 CAAGCGCCAA CAGCAGCGAC TGACTACCCA CTGACTTAGC TCCCTATTTA AAGGCACCTT  
34741 AACTGACGT AATGACCAA GGTCTAAAAA CCCC GCCAAA AAAACACACA CGCCCTGGGT  
34801 GTTTTTCGA AAACACTTCC GCGTTCTCAC TTCCTCGTAT CGATTTTCGT ACTTGACTTC  
34861 CGGGTTCCCA CGTTACGTCA CTTTGGCCCT TACATGTAAC TTAGTCGTAG GCGGCCATCT  
34921 TGCCACGTC CAAAATGGCT TACATGTCCA GTTACGCCTC CGCGGCGACC GTTAGCCGTG  
34981 CGTCGTGACG TCATTTGCAT CAACGTTTCT CGGCCAATCA GCAGTAGCCC CGCCCTAAAT  
35041 TTAAACCTC ATTTGCATAT TAACTTTTGT TTACTTTGTG GGGTATATTA TTGATGATG



ATGTCAAAGAGGCTCCGGGTGGAAGATGACTTCAACCCCGTCTACCCCTA  
TGGCTACGCGCGGAATCAGAATATCCCCTTCCTCACTCCCCCCTTTGTCTC  
CTCCGATGGATTCAAAAACCTTCCCCCCTGGGGTCTGTCACTCAAACCTGGC  
TGACCCAATCACCATAGCCAATGGTGATGTCTCACTCAAGGTGGGAGGGG  
GACTTACTTTGCAAGAAGGAAGTCTGACTGTAGACCCTAAGGCTCCCTTG  
CAACTTGCAAACAATAAAAAACTTGAGCTTGTTTATGTTGATCCATTTGAG  
GTTAGTGCCAATAAACTTAGTTTAAAAGTAGGACATGGATTAAAAATATT  
AGATGACAAAAGTGCTGGAGGGTTGAAAGATTTAATTGGCAAACCTTGTTGG  
TTTTAACAGGGAAAGGAATAGGCACTGAAAATTTGCAAATAACAGATGGT  
AGCAGCAGAGGAATTGGTATAAGTGTAAGAGCAAGAGAAGGGTTAACAT  
TTGACAATGATGGATACTTGGTAGCATGGAACCCAAAGTATGACACGCGC  
ACACTTTGGACAACACCAGACACATCTCCTAATTGCAGGATTGATAAGGA  
GAAGGATTCAAACTCACTTTGGTACTTACAAAGTGTGGAAGTCAAATAT  
TAGCTAATGTGTCTTTGATTGTGGTGTGTCAGGAAAATATCAATACATAGACC  
ACGCTACAAATCCAACCTCTTAAATCATTTAAAATAAACTTCTTTTTGATA  
ATAAAGGTGTACTTCTCCCAAGTTCAAACCTTGATTCCACATATTGGAACCT  
TTAGAAGTGACAATTTAACTGTATCTGAGGCATATAAAAATGCAGTTGAA  
TTTATGCCTAATTTGGTAGCCTACCCAAAACCTACCACTGGCTCTAAAAAA  
TATGCAAGGGATATAGTCTATGGGAACATATATCTTGGAGGTTTGGCATA  
TCAGCCAGTTGTAATTAAGGTTACTTTTAATGAAGAAGCAGATAGTGCTTA  
CTCTATAACATTTGAATTTGTATGGAATAAAGAATATGCCAGGGGGTTGAA  
TTTGAAACCACTTCCTTTACCTTCTCCTATATTGCCCAACAATAA

SEQ ID NO:2

**SUBSTITUTE SHEET (RULE 26)**

Penton17.Seq Length: 1554

1 ATGAGGCGTG CGGTGGTGTC TTCCTCTCCT CCTCCCTCGT ACGAGAGCGT  
51 GATGGCGCAG GCGACCCTGG AGGTTCCGTT TGTGCCTCCG CGGTATATGG  
101 CTCCTACGGA GGGCAGAAAC AGCATTCGTT ACTCGGAGCT GGCTCCGTTG  
151 TACGACACCA CTCGCGTGTA CTTGGTGGAC AACAAGTCGG CGGACATCGC  
201 TTCCCTGAAC TATCAAAACG ACCACAGCAA CTTCTTGACC ACGGTGGTGC  
251 AGAACAACGA TTTCACCCCC GCCGAGGCTA GCACGCAGAC GATAAATTTT  
301 GACGAGCGGT CGCGGTGGGG CGGTGATCTG AAGACCATTG TGCACACCAA  
351 CATGCCCAAT GTGAACGAGT ACATGTTTAC CAGCAAGTTT AAGGCGCGGG  
401 TGATGGTGGC TAGAAAACAC CCACAGGGGG TAGAAGCAAC AGATTTAAGC  
451 AAGGATATCT TAGAGTATGA GTGGTTTGAG TTTACCCTGC CCGAGGGCAA  
501 CTTTTCGAG ACCATGACCA TAGACCTGAT GAACAACGCC ATCTTGAAA  
551 ACTACTTGCA AGTGGGGCGG CAAAATGGCG TGCTGGAGAG CGATATTGGA  
601 GTCAAGTTTG ACAGCAGAAA TTTCAAGCTG GGCTGGGACC CTGTGACCAA  
651 GCTGGTGATG CCAGGGGTCT ACACCTACGA GGCCTTTCAC CCGGACGTGG  
701 TGCTGCTGCC GGGCTGCGGG GTGGACTTCA CAGAGAGCCG CCTGAGCAAC  
751 CTCCTGGGCA TTCGCAAGAA GCAACCTTTC CAAGAGGGCT TCAGAATCAT  
801 GTATGAGGAT CTAGAAGGGG GCAACATCCC CGCCCTGCTG GATGTGCCCCA  
851 AGTACTTGGA AAGCAAGAAG AAGTTAGAGG AGGCATTGGA GAATGCTGCT  
901 AAAGCTAATG GTCCTGCAAG AGGAGACAGT AGCGTCTCAA GAGAGGTTGA  
951 AAAGGCAGCT GAAAAAGAAC TTGTTATTGA GCCCATCAAG CAAGATGATA  
1001 CCAAGAGAAG TTACAACCTC ATCGAGGGAA CCATGGACAC GCTGTACCGC  
1051 AGCTGGTACC TGTCTATAC CTACCGGGAC CCTGAGAACG GGGTGCAGTC  
1101 GTGGACGCTG CTCACCACCC CGGACGTAC CTGCGGCGCG GAGCAAGTCT  
1151 ACTGGTCGCT GCCGGACCTC ATGCAAGACC CCGTCACCTT CCGTTCTACC  
1201 CAGCAAGTCA GCAACTACCC CGTGGTCGGC GCCGAGCTCA TGCCCTTCCG  
1251 CGCCAAGAGC TTTTACAACG ACCTCGCCGT CTACTCCCAG CTCATCCGCA  
1301 GCTACACCTC CCTCACCCAC GTCTTCAACC GCTTCCCCGA CAACCAGATC

SEQ ID NO: 3

1351 CTCTGCCGTC CGCCCGCGCC CACCATCACC ACCGTCAGTG AAAACGTGCC  
1401 TGCTCTCACA GATCACGGGA CGCTACCGCT GCGCAGCAGT ATCCGCGGAG  
1451 TCCAGCGAGT GACCGTCACT GACGCCCCGTC GCCGCACCTG TCCCTACGTC  
1501 TACAAGGCCC TGGGCATAGT CGCGCCGCGT GTGCTTTCCA GTCGCACCTT  
1551 CTAA



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Claims

1. A chimeric adenoviral vector comprising nucleotide sequence of a first adenovirus, wherein at least one gene of said first adenovirus encoding a protein that facilitates binding of said vector to a target mammalian cell, or internalization thereof within said cell, is replaced by the corresponding gene from a second adenovirus belonging to subgroup D, said vector further comprising a transgene operably linked to a eucaryotic promoter to allow for expression therefrom in a mammalian cell.
2. A chimeric adenoviral vector according to Claim 1 wherein said second adenovirus is selected from the group consisting of Ad 9, Ad 15, Ad 17, Ad 19, Ad 20, Ad 22, Ad 26, Ad 27, Ad 28, Ad 30, and Ad 39.
3. A chimeric adenoviral vector according to Claim 1 wherein said first adenovirus is selected from the group consisting of Ad 2, Ad 5, and Ad 12.
4. A chimeric adenoviral vector according to Claim 1 wherein said replaced gene encodes Ad fiber.
5. A chimeric adenoviral vector according to Claim 1 wherein said replaced gene encodes Ad penton base.
6. A chimeric adenoviral vector according to Claim 1 wherein a first replaced gene encodes Ad fiber, and a second replaced gene encodes Ad penton base.
7. A chimeric adenoviral vector comprising nucleotide sequence of a first adenovirus, wherein a portion of a gene thereof encoding a protein that facilitates binding of said vector to a target mammalian cell, or internalization

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thereof within said cell, is replaced by a portion of the corresponding gene from a second adenovirus belonging to subgroup D, said vector further comprising a transgene operably linked to a eucaryotic promoter to allow for expression therefrom in a mammalian cell.

5

8. A chimeric adenoviral vector according to Claim 7 wherein the encoding sequence that is replaced codes for a portion of Ad fiber.

9. A chimeric adenoviral vector according to Claim 7 wherein the encoding  
10 sequence that is replaced codes for a portion of Ad penton base.

10. A chimeric adenoviral vector according to Claim 9 wherein the encoding sequence that is replaced codes for an amino acid sequence that includes RGD.

15 11. A method of providing a biologically active protein to the airway epithelial cells of a patient comprising administering to said cells an adenoviral vector selected from the group consisting of:

(a) a chimeric adenoviral vector comprising nucleotide sequence of a first adenovirus, wherein at least one gene of said first adenovirus  
20 encodes a protein that facilitates binding of said vector to a target mammalian cell, or internalization thereof within said cell, is replaced by the corresponding gene from a second adenovirus belonging to subgroup D, said vector further comprising a transgene encoding said protein that is operably linked to a eucaryotic promoter to allow for  
25 expression therefrom in a mammalian cell; and

(b) a chimeric adenoviral vector comprising nucleotide sequence of a first adenovirus, wherein a portion of a gene thereof encoding a protein that facilitates binding of said vector to a target mammalian cell, or internalization thereof within said cell, is replaced by a portion of the

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corresponding gene from a second adenovirus belonging to subgroup D, said vector further comprising a transgene encoding said protein that is operably linked to a eucaryotic promoter to allow for expression therefrom in a mammalian cell;

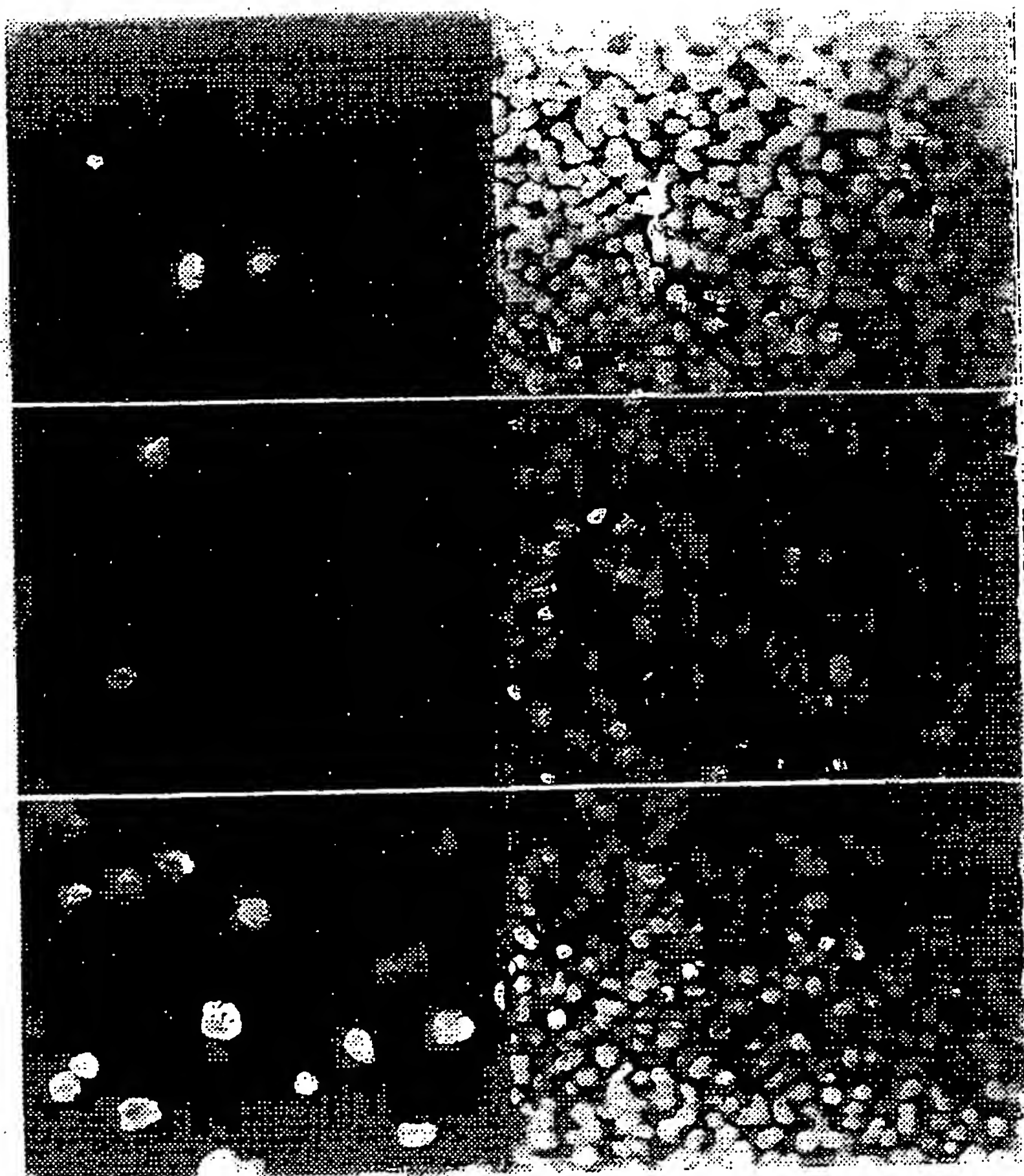
5 under conditions whereby the transgene encoding said protein is expressed, and phenotypic benefit is produced in said airway epithelial cells.

12. A method according to Claim 11 wherein said second adenovirus is Ad 17 and the nucleotide sequence thereof used in replacement of nucleotide sequence of  
10 said first adenovirus encodes a polypeptide selected from the group consisting of Ad 17 fiber, a fragment of Ad 17 fiber, Ad 17 hexon, a fragment of Ad 17 hexon, Ad penton base, and a fragment of Ad 17 penton base.

13. A method of providing a biologically active protein to the airway epithelial  
15 cells of a patient that comprises administering to said cells an adenoviral vector comprising elements of an Ad 17 genome, and a transgene encoding said protein that is operably linked to a eucaryotic promoter to allow for expression therefrom in a mammalian cell, under conditions whereby the transgene encoding said protein is expressed, and phenotypic benefit is  
20 produced in said airway epithelial cells.

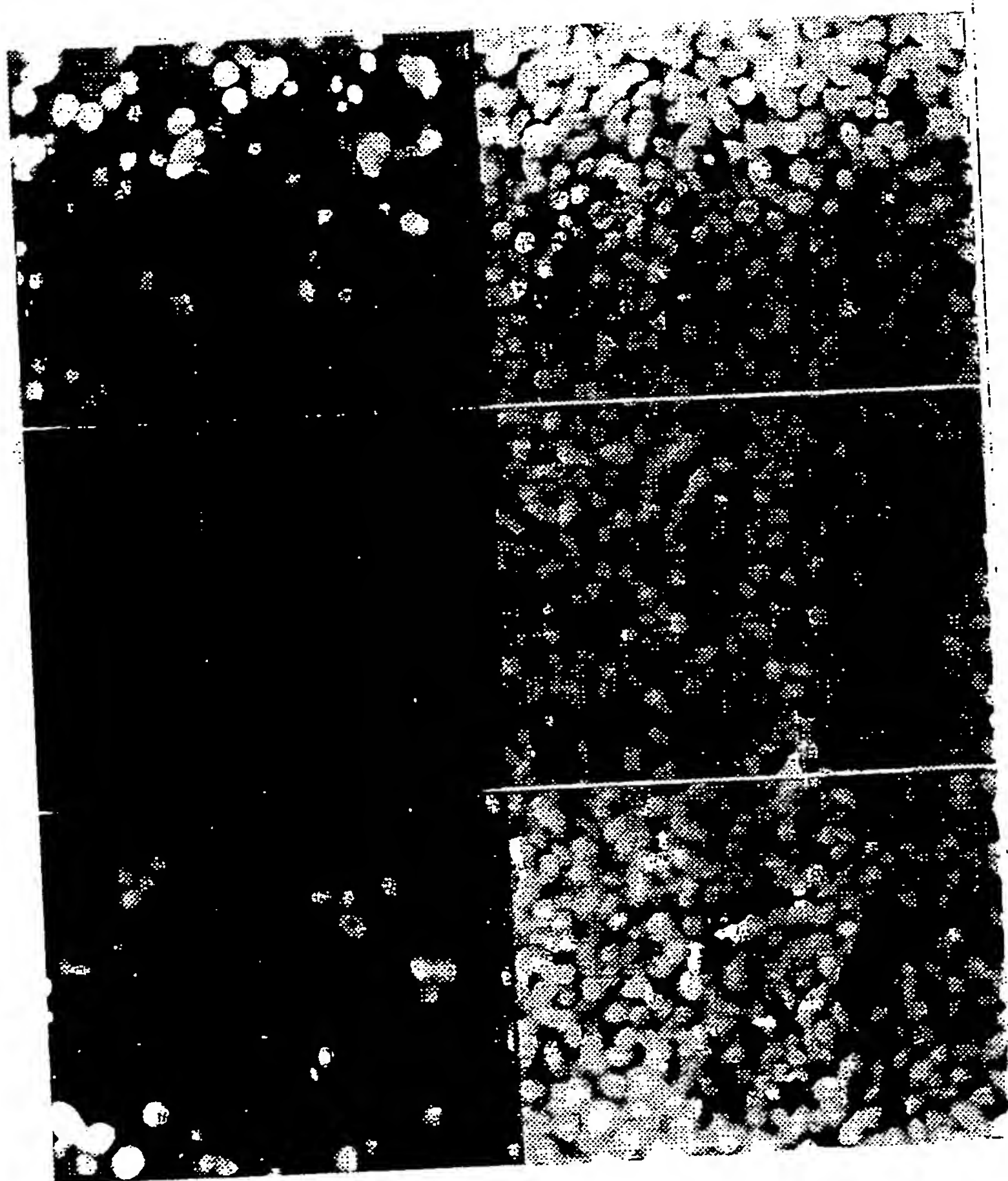
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*FIG. 1*



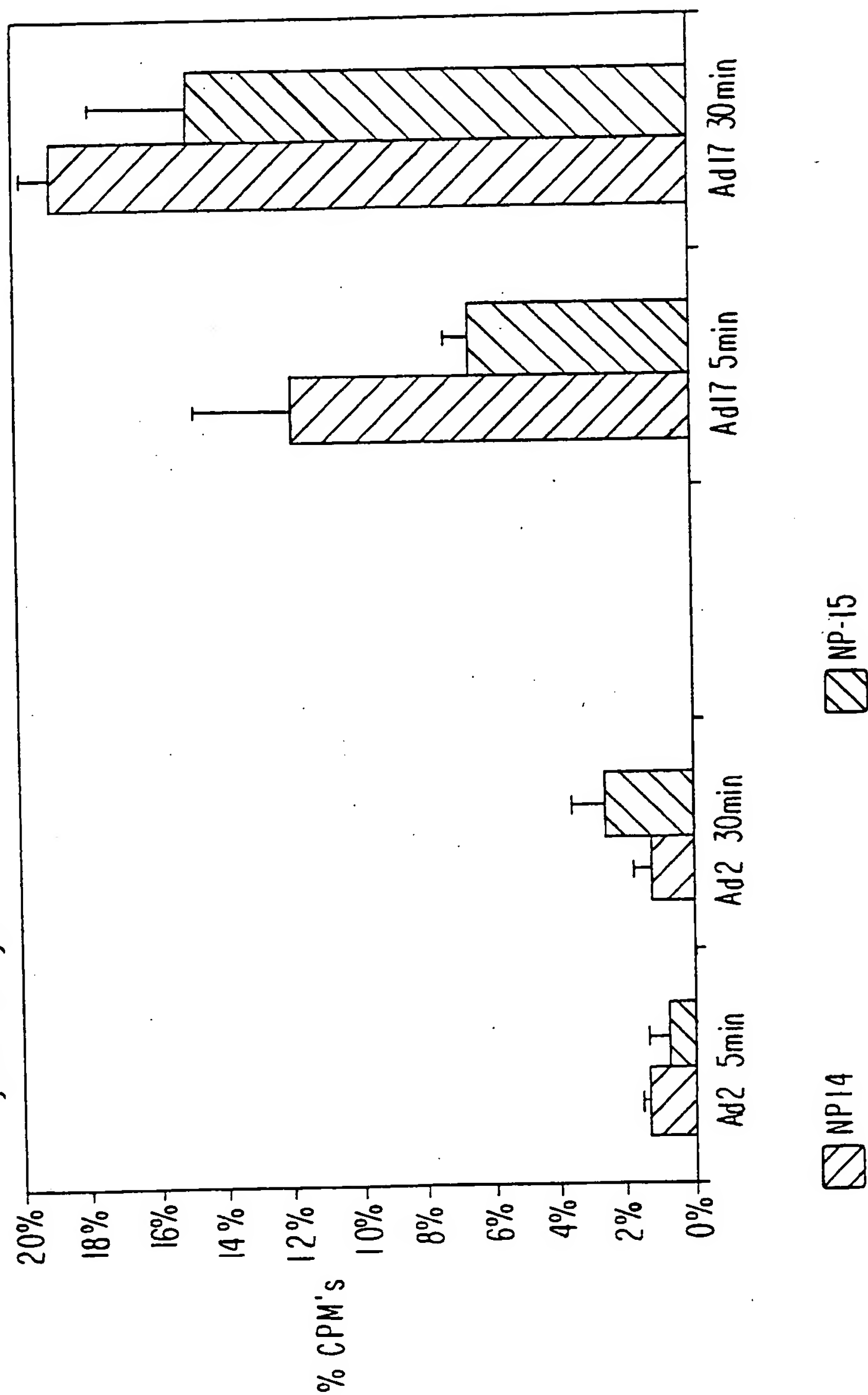
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*FIG. 2*



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**FIG. 3**  
Binding to Ciliated Human Airway Epithelia  
by Ad2 and by Ad17



# Chimeric Ad2/βgal-2/ Ad17 vectors

**FIG. 4**

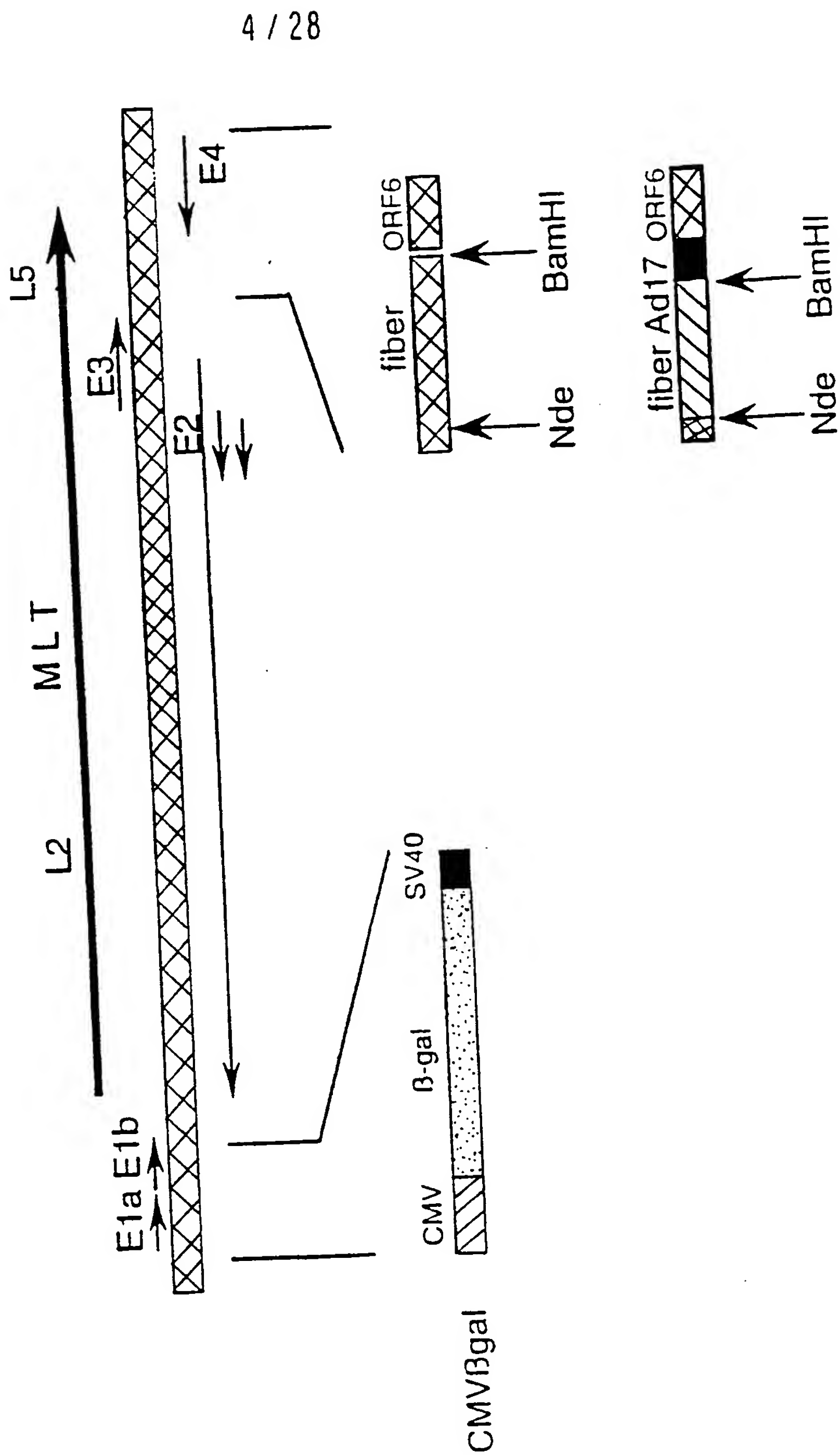






FIG. 5A-2

```

START_____
      |
240 GVDFTESRLSNLLGIRKKQPFQEGFRIMYEDLEGGNIPALLDVPKYLES. 288
    |||||.|||||:|||||:|||||:|||||.|||.
247 GVDFTHSRLSNLLGIRKRQPFQEGFRITYDDLEGGNIPALLDVDAYQASL 296
      .
      .RGDSSVS 313
289 KKKLEEALENA AKANGPA.....
    |..|:::|:|...:
297 KDDTEQG GAGGNNSCSGAEENSNA AAMQPVEDMNDHAI RGDTFAT 346
      .
      .KELVIEPIKQDDTKRSYNLIEG 343
314 REVEKAAE.....
    |.|||||
347 RAEEKRAEA EAAEAAPAAQPEVEKPQKKPV IKPLTEDSKKRSYNLISN 396
      .
      .TLYRSWYLSYTYRDPENG VQSWTL TTPDVTCGA EQVWSLPDLMQ 392
344 TMD.TLYRSWYLSYTYRDPENG VQSWTL TTPDVTCGA EQVWSLPDLMQ 446
      .
397 DSTFTQYRSWYLAYNYGDPQTGI RSWTLLCTPDVTCGSEQVWSLPDMMQ
      |
END

```

FIG. 5B

393 DPVTFRSTQQVSNYPVVGAEELMPFRAKSFYNDLAVYSQLIRSYTSLTHVF 442  
 ||||| | : || : ||||| : | : ||||| : |||||  
 447 DPVTFRSTQISNFPVVGAEELPVHKSIFYNDQAVYSQLIRQFTSLTHVF 496  
 443 NRFPDNQILCRPPAPTITTVSENVPAALTDHGTLPLRSSIRGVQRTVTDA 492  
 |||| : |||| . ||||| ||||| . ||||| : ||||  
 497 NRFPENQILARPAPTITTVSENVPAALTDHGTLPLRNSIGGVQRTITDA 546  
 493 RRRTCPVYKALGIVAPRVLSSRTF 517  
 ||||| ||||| . ||||| |||||  
 547 RRRTCPVYKALGIVSPRVLSSRTF 571

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FIG. 6A-1

	1	50	
Penton5	1	SPFDAPLDPP~SEQ ID NO:6	APVAAALG
Penton2	...	SPFDAPLDPP~SEQ ID NO:5	PPSYESVUSA
Penton3	...	AMIQPPLEAP~SEQ ID NO:7	PPSYESVUSA
Penton12	...	AAAPP~SEQ ID NO:8	PPSYESVM..
Penton40	...	ADLPATLQAL~SEQ ID NO:9	PPSYETVM..
Penton17	...	QATLEVP~SEQ ID NO:4	PPSYESVM..
Pentonf10	...	CSSSGGQSEL~SEQ ID NO:10	PPSYESVM..
	MWGLQPPTSI	YPPPAASAQS	PSTYPAMVNG
	51	VASLNYQNDH	ELAPLFDTTR
Penton5	FVP.PRYLRP	VASLNYQNDH	ELAPLFDTTR
Penton2	FVP.PRYLRP	IASLNYQNDH	DVSPLYDTTK
Penton3	FVP.PRYLAP	IASLNYQNDH	ELSPLYDTTR
Penton12	YVP.PRYLGP	IASLNYQNDH	ELAPLYDTTR
Penton40	HVP.PRYLGP	IASLNYQNDH	ELAPLYDTTR
Penton17	FVP.PRYMAP	IDTYNKKDANH	DYTPCRNTTK
Pentonf10	YMPLQRVMAP		

FIG. 6A-2

					150
	101				
Penton5	SNFLTTVIQN	NDYSPGEAST	QTINLDDRRSH	WGGDLKTIH	TNMPNVNEFM
Penton2	SNFLTTVIQN	NDYSPGEAST	QTINLDDRRSH	WGGDLKTIH	TNMPNVNEFM
Penton3	SNFLTTVVQN	NDFTPTEAST	QTINFDESR	WGGQLKTIH	TNMPNVNEYM
Penton12	SNFLTTVVQN	NDYSPIEAGT	QTINFDESR	WGGDLKTIH	TNMPNVNDFM
Penton40	SNFQTTVVQN	NDFTPTEAGT	QTINFDDRRS	WGGDLKTIH	TNMPNINEFM
Penton17	SNFLTTVVQN	NDFTPAEAST	QTINFDESR	WGGDLKTIH	TNMPNVNEYM
Pentonf10	SNFRTTVIHN	QDLADTAAT	ESIQLDNRSC	WGGDLKTAVR	TNCPNVSSFF

					200	9/28
	151					
Penton5	FTNKFKARVM	VSRL.....	...PTKD..N	QVELKYEWVE	FTLPEGNYSE	
Penton2	FTNKFKARVM	VSRS.....	...LTKD..K	QVELKYEWVE	FTLPEGNYSE	
Penton3	FSNKFKARVM	VSRLAPEGVT	VNDTYDH..K	EDILKYEWFE	FILPEGNFSA	
Penton12	FTTKFKARVM	VARK.....	...TNNE..G	QTILEYEWA	FVLPEGNYSE	
Penton40	STNKFRARVM	VEK.....	...VNR..K	TNAPRYEWFE	FTLPEGNYSE	
Penton17	FTSKFKARVM	VARKHPOGV.	...EATDL..S	KDILEYEWFE	FTLPEGNFSE	
Pentonf10	QNSVVRVRMM	WKRDPPTSTA	PPSAVSGSYS	VPGAQYKWD	LTVPEGNYAL	

					250
	201				
Penton5	TMTIDLNMNA	IVEHYLKVGR	QNGVLESDIG	VKFDTRNFRL	GFDPVTGLVM

FIG. 6B-1

Penton2	TMTIDLMNNA	IVEHYLKVGR	QNGVLESDIG	VKFDTRNFRL	GFDPVTGLVM
Penton3	TMTIDLMNNA	IIDNYLEIGR	QNGVLESDIG	VKFDTRNFRL	GWDPETKLIM
Penton12	TMTIDLMNNA	IIEHYLRVGR	QHGVLSDIG	VKFDTRNFRL	GWDPETQLVT
Penton40	TMTIDLMNNA	IVDNYLAVGR	QNGVLESDIG	VKFDTRNFRL	GWDPVTKLVM
Penton17	TMTIDLMNNA	I LENYLQVGR	QNGVLESDIG	VKFDSRNFKL	GWDPVTKLVM
Pentonf10	CELIDLLNEG	IVQLYLSEGR	QNNVQKSDIG	VKFDTRNFGL	LRDPVTGLVT
					300
Penton5	PGVYTNEAFH	PDIILLPGCG	VDFTHSRLSN	LLGIRKRQPF	QEGFRITYDD
Penton2	PGVYTNEAFH	PDIILLPGCG	VDFTHSRLSN	LLGIRKRQPF	QEGFRITYDD
Penton3	PGVYTYEAFH	PDIVLLPGCG	VDFTESRLSN	LLGIRKRHPF	QEGFKIMYED
Penton12	PGVYTNEAFH	PDIVLLPGCG	VDFTESRLSN	ILGIRKRQPF	QEGFVIMYEH
Penton40	PGVYTNEAFH	PDIVLLPGCG	VDFTSRLNN	LLGIRKRMPF	QKGFQIMYED
Penton17	PGVYTYEAFH	PDVVLPGCG	VDFTESRLSN	LLGIRKKQPF	QEGFRIMYED
Pentonf10	PGTYVYKGYH	PDIVLLPGCA	IDFTYSRLSL	LLGIGKREPY	SKGFVITYED



# FIG. 6B-2

	301					350
Penton5	LEGGNIPALL	DVDAYQASLK	DDTEQGGGA	GGNSSGSGA	EENSNAATAA	
Penton2	LEGGNIPALL	DVDAYQASLK	DDTEQGGGA	GGNSSGSGA	EENSNAATAA	
Penton3	LEGGNIPALL	DVTAYEESKK	DTTETTTLA	VAEETSE...	.....	
Penton12	LEGGNIPALL	DVKKYENSL.	.....	.....Q...	.....	
Penton40	LEGGNIPALL	DVEKYEASIK	.....	.....	.....	
Penton17	LEGGNIPALL	DVPKYLESKK	KLE.....E	ALENAAK...	.....	
Pentonf10	LQGGDIPALL	DLDSVDVND	A.....	DGEVIELDNA	A.....	
	351					400
Penton5	MQPVEDMNDH	AIRGDTFATR	AEEKRAEAE	AAEAAAPAAQ	PEVEKPQKKP	
Penton2	MQPVEDMNDH	AIRGDTFATR	AEEKRAEAE	AAEAAAPAAQ	PEVEKPQKKP	
Penton3	.....DDD	ITRGDTYITE	KQKREAAAE	V.....	.....KKEL	
Penton12	.....DQN	TVRGDNFIA.	.....	L.....	.....NKAA	
Penton40	.....EAQ	EIRGADEFKPN	PQ.....	.....	.....DL	
Penton17	.....ANG	PARGDSSVSR	EVEKAA...	.....	.....EKEL	
Pentonf10	.....	.....	.....	.....	.....	

FIG. 6B-3

450

401	VIKPLTEDSK	KRSYNLI...	SNDSTFTQYR	SWYLAYNYGD	PQTGIRSWTL
penton5	VIKPLTEDSK	KRSYNLI...	SNDSTFTQYR	SWYLAYNYGD	PQTGIRSWTL
penton2	KIQPLEKDSK	SRSYNVL...	E.DKINTAYR	SWYLSYNYGN	PEKGIRSWTL
penton3	RIEPVETDPK	GRSYNLL...	P.DKKNTKYR	SWYLAYNYGD	PEKGVRSWTL
penton12	EIVPVEKDSK	ERSYNLL...	EGDKNNTAYR	SWFLAYNYGD	AEKGVKSWTL
penton40	VIEPIKQDDT	KRSYNLI...	E.GTMDTLYR	SWYLSYTYRD	PENGVOQSWTL
penton17	... PLLHDSA	GVSYNVIYDQ	VTGKPVTAIR	SWMLAYNVPN	SQANQT...TL
pentonf10					

500

451		LCTPDVTCGS	EQVYWSLPDM	MQDPVTFRST	RQISNFPVVG	AELLPVHSKS
Penton5		LCTPDVTCGS	EQVYWSLPDM	MQDPVTFRST	SQISNFPVVG	AELLPVHSKS
Penton2		LCTPDVTCGS	EQVYWSLPDM	MQDPVTFRST	RQVNNYPVVG	AELMPVFSKS
Penton3		LTTSADVTCGA	EQVYWSLPDM	MQDPVTFRSS	RQVSNYPVVA	AELLPVHAKS
Penton12		LTTPDVTCGS	EQVYWSLPDM	MQDPVTFRPS	TQVSNYPVVG	VELLPVHAKS
Penton40		LTtTDVTCGS	QQVYWSLPDM	MQDPVTFRST	QQVSNYPVVG	AELMPFRAKS
Penton17		LTTPDVTCGA	EQVYWSLPDL	MQDPVTFRST	NTTNLCPVVG	MNLFPTYNKI
Pentonf10		LTVPDMAGGI	GAMYTSLPDT	FIAPTGFKE		

550

501	FYNDQAVYSQ	LIRQFT.SLT	HVFNRFPENQ	ILARPPAPTI	TTVSENVVPAL
Penton5	FYNDQAVYSQ	LIRQFT.SLT	HVFNRFPENQ	ILARPPAPTI	TTVSENVVPAL
Penton2	FYNDQAVYSQ	LIRQFT.SLT	HVFNRFPENQ	ILIRPPAPTI	TTVSENVVPAL
Penton3	FYNEQAVYSQ	QLRQAT.SLT	HVFNRFPENQ	ILVRPPAATI	TTVSENVVPAL
Penton12	FYNEQAVYSQ	LIRQST.ALT	RVFNRFPENQ		

FIG. 6C

Penton40	FYNEQAVYSQ	LIRST.ALT	HIFNRFPENQ	ILVRPPAPTI	TTVSENVPAL
Penton17	FYNDLAVYSQ	LIRSYT.SLT	HVFNRFPDNQ	ILCRPPAPTI	TTVSENVPAL
Pentonf10	YYQAASTYVQ	RENSCQSAT	AEFNRFPENE	ILKQAPPMNV	SSVCDNQPAV
	551				600
Penton5	TDHGTLPLRN	SIGGVQRVTI	TDARRRTCPI	VYKALGIVSP	RVLSSRTF**
Penton2	TDHGTLPLRN	SIGGVQRVTI	TDARRRTCPI	VYKALGIVSP	RVLSSRTF**
Penton3	TDHGTLPLRS	SIRGVQRVTI	TDARRRTCPI	VYKALGIVAP	RVLSSRTF**
Penton12	TDHGTLPLRS	SISGVQRVTI	TDARRRTCPI	VYKALGIVSP	RVLSSRTF**
Penton40	TDHGTLPLRS	SISGVQRVTI	TDARRRTCPI	VHKALGIVAP	KVLSSRTF**
Penton17	TDHGTLPLRS	SIRGVQRVTI	TDARRRTCPI	VYKALGIVAP	RVLSSRTF**
Pentonf10	VQQGVLPVKS	SLPGLQRVLI	TDDQRRPIPI	VYKSIATVQP	TVLSSATLQ*

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FIG. 7A-1

Fiber17.pep x Fiber2.pep

[illegible]

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## FIG. 7A-2

145 .....NLQNTD...GSSRGIGISVRARE 164  
|::||. ....|:::|:::.....|  
301 YNRGLYLEFNASNNTKKLEVSIKKSSGLNFDNTAIAINAGKGLEFDTNTSE 350  
165 .....GLTFDNDGYLVAWNPKYDTRT 185  
||.||||. |::|::|::|::|  
351 SPDINPIKIGSGIDYNEENGAMITKLGAGLSFDNSGAITIGNKNDKLT 400  
186 LWTPDTPSPNCRIDKEKDSKLTVLTKCGSQILANVSLIVVSGKYQYIDH 235  
|||||.|||||.:::|:::|||||:::|:::|:::|  
401 LWTPDTPSPNCRIHSDNDCKFTLVLTCKGSQVLATVAALAVSGDLS.... 446

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FIG. 7B

236 ATNPTLKSEFKIKLLFDNKGVLPPSSNLDSTYWNFRSDNLTVSEAYKNAVE 285  
 447 SMTGTVASVSI FLRFDQNGVLMENSS LKKHYWNFRNGNSTNANPYTNAV G 496  
 286 FMPNLVAYPKPTTGSKKKYARDIVYGN IYLGGLAYQP VVIKVT FNEEAD.. 333  
 497 FMPNLLAYPKTQSQT... AKNNIVSQVYLHGDKTKPMIL TITLNGTSEST 543  
 334 .....SAYSITFEFVWNKE..YARVEFETSTFTFSYIAQQ 366  
 544 ETSEVSTYSMSFTWSWESGKYTTETFATNSYTF SYIAQE 582



FIG. 8A-1

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8fiber	1	MTKRLRA...	...	EDDFN	PVYPYGYARN	Q.NIPFLTPP	FVSSNGFQNF	50
9fiber		MSKRLRV...	...	EDDFN	PVYPYGYARN	Q.NIPFLTPP	FVSSDGFQNF	
15fiber		MSKRLRV...	...	EDDFN	PVYPYGYARN	Q.NIPFLTPP	FVSSDGFQNF	
17fiber		MSKRLRV...	...	EDDFN	PVYPYGYARN	Q.NIPFLTPP	FVSSDGFQNF	
2fiber		.MKRARP...	...	SEDTFN	PVYPYDTETG	PPTVPFLTPP	FVSPNGFQES	
5fiber		.MKRARP...	...	SEDTFN	PVYPYDTETG	PPTVPFLTPP	FVSPNGFQES	
4fiber		MSKSARG...	...	WSDGFD	PVYPYDADND	RP.CPSSTLP	SFSSDGFQEK	
40-1fiber		.MKRTRIE...	...	DDFN	PVYPYD.TSS	TPSIPYVAPP	FVSSDGLQEN	
41fiber		.MKRTRIE...	...	DDFN	PVYPYD.TFS	TPSIPYVAPP	FVSSDGLQEK	
40-2fiber		.MKRARFE...	...	DDFN	PVYPYE.HYN	PLDIPFITPP	FASSNGLQEK	
12fiber		.MKRSRTQYA	EETEENDDFN		PVYPFD.PFD	TSDVPFVTPP	FTSSNGLQEK	
3fiber		MAKRARL...	...	STSFN	PVYPYEDESS	SQH.PFINPG	FISPDGFTQS	

FIG. 8A-2

[illegible]

FIG. 8B-1

2fiber	KTKSNISLDT	FAPLTIITSGA	LTVATTAPLI	VTSGALSVQS	QAPLTVQDSK
5fiber	KTKSNINLEI	SAPLTVTSEA	LTVAAPAPLM	VAGNTLTMQS	QAPLTVHDSK
4fiber	.....	.....	.....	SFFQQH	HFPL.....
40-1fiber	.....	.....	.....	.....	.....
41fiber	.....	.....	.....	.....	.....
40-2fiber	KTNKIVGLNY	TKPLALQNNA	LTLSYNAPFN	VVNNNLALNM	SQPVTI.....
12fiber	NTSQGLKLSW	SAPLAVKASA	LTLNTRAPLT	TTDESLALIT	APPITVESSR
3fiber	.....	.....	.....	.....	.....
	151				200
8fiber	.....	.....	.....	.....	.....
9fiber	.....	.....	.....	.....	.....
15fiber	.....	.....	.....	.....	.....
17fiber	.....	.....	.....	.....	.....
2fiber	.....	.....	.....	.....	LSI
5fiber	.....	.....	.....	.....	LSI
4fiber	.....	.....	.....	.....	.....
40-1fiber	.....	.....	.....	.....	.....
41fiber	.....	.....	.....	.....	.....
40-2fiber	.....	NANNELSLI	IDAPLNADTG	TLRLRSDAPL	GLVDK.TLKV
12fiber	LGLATIAPLS	LDGGGNLGLN	LSAPLDVSNN	NLHLTTETPL	VVNSSGALSV
3fiber	.....	.....	.....	.....	.....

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FIG. 8B-2

	201	GKLT	VNTEPPLH..	.....	250
8fiber	.....	.....	VNADPPLQ..	.....	
9fiber	.....	.....	VNTEPPLQ..	.....	
15fiber	.....	.....	VDPKAPLQ..	.....	
17fiber	.....	.....	VTASPPLTTA	TGSLGINMED	
2fiber	ATKGPI TVSD	GKLALQTSAP	ITASPPLTTA	TGSLGIDLKE	2/28
5fiber	ATQGPLTVSE	GKLALQTSQP	YKFLPPLSIL	KSTI.....	
4fiber	.....	.....TWIP	..VSPPLTNS	NNSLGLATSA	
40-1fiber	.....	...TVPT...	..VSPPLTNS	NNSLGLATSA	
41fiber	.....	...TVPT...	LKYSPPLKIE	NENLTLSTGG	
40-2fiber	LFSSPLYLDN	NFLT LAIERP	ISVTSPTVI	NGSLALSTTA	
12fiber	ATADPISVRN	NALTLP TADP	.....	.....	
3fiber	.....	.....	.....	.....	

FIG. 8B-3

8fiber	251	..LTNN.KLG	IALDAPFDVI	D..NKLTLLA	GHGLSII.TK	ETSTLPGLVN	300
9fiber		..LTNN.KLG	IALDAPFDVI	D..NKLTLLA	GHGLSII.TK	ETSTLPGLRN	
15fiber		..LTNN.RIG	IALDAPFDVI	G..GKLTLLA	GHGLSII.TE	ETSPLPGLVN	
17fiber		..LANNKKLE	LVYVDPFEVS	A..NKLSLKV	GHGLKILDDK	SAGGLKDLIG	
2fiber		PIYVNNKGIG	IKISGPLQVA	QNSDTLTVVT	GPGVTVEQNS	LRTKVAGAIG	
5fiber		PIYTQNGKLG	LKYGAPLHVT	DDLNTLTVAT	GPGVTINNNTS	LQTKVTGALG	
4fiber		.....	.....	..LNTLVSAF	GSGLGLSGSA	LAVQLASPLT	21/28
40-1fiber		PIAVSANSALT	LATAAPLTVS	N..NQLSINT	GRGLVITNNA	VAVNPTGALG	
41fiber		PIAVSANSALT	LATAAPLTVS	N..NQLSINA	GRGLVITNNA	LTVNPTGALG	
40-2fiber		PFTVSGGNLN	LATSAPLSVQ	N..NSLSLGV	NPPFLITDSG	LAMDLDGDLA	
12fiber		PLNSTGSTLS	LSVANPLTIS	Q..DTLTVST	GNGLQVSGSQ	LVTRIGDGLT	
3fiber		...TTDGSLE	ENIKVNTPLT	KSNHSINLPI	GNGLQIEQNK	LCS.....	
		301				350	
8fiber		.....	.....	.....	.....	.....	
9fiber		.....	.....	.....	.....	.....	
15fiber		.....	.....	.....	.....	.....	
17fiber		.....	.....	.....	.....	.....	
2fiber		YDSSNNMEIK	TGGMRIN..	NNLLILDVDY	PFDAQTKLRL	KLGGQPLYIN	

FIG. 8C-1

5fiber	FDSQGNMQLN	VAGGLRIDSQ	NRRLLDVSQ	PFDAQNQLNL	RLGQGPLFIN
4fiber	FDDKG				
40-1fiber	FNNTGALQLN	AAGGMRVDGA	N..LILHVAY	PFEAINQLTL	R.....
41fiber	FNNTGALQLN	AAGGMRVDGA	N..LILHVAY	PFEAINQLTL	R.....Q
40-2fiber	LGG.SKLIIN	LGPGQLQMSNG	A..ITL.....	ALDAALPL..	RRGLGLIYNQ
12fiber	FDN.GVMKVN	VAGGMRTSGG	R..IILDVNY	PFDASNNLSL	
3fiber					
	351				22/28 400
8fiber					TLVVLTKGKI
9fiber					TLVVLTKGKI
15fiber					TLVVLTKGKL
17fiber					KLVLTKGKI
2fiber	ASHNLDINYN	RGLYLEFNASN	NTKKLEVSIK	KSSGLNFDNT	AIAINAGKGL
5fiber	SAHNLDINYN	KGLYLEFTASN	NSKKLEVNLS	TAKGLMFDAT	AIAINAGDGL
4fiber	..NIKITLN	RGLHVTTGDA	..IESNIS	WAKGIKFEDG	AIAINAGKGL
40-1fiber					
41fiber					QLQLRIGS
40-2fiber	YKNN				NLTTDIST
12fiber	STNW				
3fiber					



FIG. 8C-2

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8 fiber	401	GTDLNNGG.	...	NICVRVG	E.....	...	GGGLS	FNDNGDLVAF	450
9 fiber		GTESTDNNGG.	...	TVCVRVG	E.....	...	GGGLS	FNNDGDLVAF	
15 fiber		GTDTTDNNGG.	...	SIRVRVG	E.....	...	GGGLS	FNEAGDLVAF	
17 fiber		GTENLQNTDG	SSRGIGISVR	A.....	...	...	REGLT	FDNDGYLVAV	
2 fiber		EFDTNTSESP	DINPIKTKIG	SGIDYNENGA	MITKLGAGLS	FDNSGAI	TIG		
5 fiber		EFG..SPNAP	NTNPLKTKIG	HGLEFDSNKA	MVPKLGTGLS	FDSTGAI	TVG		
4 fiber		RFGTSSTETG	VNNAYPIQV.	.....	...	KLGSGLS	FDSTGAIMAG		
40-1 fiber		.....	.....	LE	NGLEVTNGGK	LNVKLGSGLQ	FDNNGRITIS		
41 fiber		.....	.....	LE	NGLEVTSGGK	LNVKLGSGLQ	FDSNGRIAIS		
40-2 fiber		ASALIMSGVT	QTLNVNANTS	KGLAIENNS.	LVVKLGNGLR	FDSWGSIAVS			
12 fiber		EKGLMFSGN.	...QIALNAG	QGLTFNNGQ.	LRVKLGAGLI	FDSNNNIALG			
3 fiber		.....	.....	.....	...	KLGNGLT	FDSSNSIALK		

FIG. 8C-3

451  
 8fiber  
 9fiber  
 15fiber  
 17fiber  
 2fiber  
 5fiber  
 4fiber  
 40-1fiber  
 41fiber  
 40-2fiber  
 12fiber  
 3fiber  
 500  
 QDKDSKLSLV LTKCGSQILA  
 QDKDSKLTLV LTKCGSQILA  
 EDKDSKLTLI LTKCGSQILG  
 KEKDSKLTLV LTKCGSQILA  
 SDNDCKFTLV LTKCGSQVLA  
 AEKDAKLTLV LTKCGSQILA  
 AENDAKLTLC LTMCD SQILA  
 ETQDANLFLC LTKNGAHVVG  
 ETQDANLFLC LTKNGAHVVG  
 ESLDAKVWLV LVKCNMVG  
 QELDAKLTLC LTKNGSIVNG  
 QNPDSKLTLI LVKNGGIVNG  
 SPNCRID...  
 SPNCKID...  
 SPNCKII...  
 SPNCRID...  
 SPNCRIH...  
 SPNCRIN...  
 SPNCQIL...  
 TPNCISIY...  
 TPNCISIY...  
 SPNATFY...  
 PPNCSLI...  
 EANCIIIEYK  
 RTLWTTTTPDT  
 RTLWTTTTPDT  
 RTLWTTTTPDP  
 RTLWTTTTPDT  
 LTLWTTTTPDP  
 LTLWTTTTPAP  
 LTLWTTTTPDP  
 LTTIWSIS.P  
 LTTIWSIS.P  
 TTLWTTTADP  
 LTLWTTTTPDP  
 TLWTGPKP  
 ALKGFTIK  
 ALKGFTIK  
 NEADKQITVK  
 TLKSFKIK  
 TGTVASVSIF  
 SGTVQSAHLI  
 LLFDKNGVLM  
 LLFDENGVLV  
 LLFDANGVLK  
 LLFDNKGVLL  
 LRFDQNGVLM  
 IRFDENGVLV  
 ESSN...  
 ESSN...  
 QGST...  
 PSSN...  
 ENSS...  
 NNSF...  
 550  
 501  
 NVSLIVVAGR  
 NVSLIVVDGK  
 SVSLLVVKGK  
 NVSLIVVSGK  
 TVAALAV.S.  
 TVSVLAV.K.  
 YKIIINNNTNP  
 YKIIINNNTQP  
 FSNINNNTNP  
 YQYIDHATNP  
 GDLSSM  
 GSLAPI  
 ALKGFTIK  
 ALKGFTIK  
 NEADKQITVK  
 TLKSFKIK  
 TGTVASVSIF  
 SGTVQSAHLI  
 LLFDKNGVLM  
 LLFDENGVLV  
 LLFDANGVLK  
 LLFDNKGVLL  
 LRFDQNGVLM  
 IRFDENGVLV  
 ESSN...  
 ESSN...  
 QGST...  
 PSSN...  
 ENSS...  
 NNSF...  
 8fiber  
 9fiber  
 15fiber  
 17fiber  
 2fiber  
 5fiber

FIG. 8D-1

4fiber	TVSVLVVRS.	....GNLNPI	TGTVSSAQVF	LRFDANGVLL	TEHS.....
40-1fiber	TITIKGLKGA	LREMNDNA..	.....LSVK	LPFDNQGNLL	NCA.....
41fiber	TITIKGLKGA	LREMHDNA..	.....LSLK	LPFDNQGNLL	NCA.....
40-2fiber	TISIKAQKGT	LL..KPTASF	.....ISFV	MYFYSDGTWR	KNYPVFDNEG
12fiber	IVSLVGVKGN	LLNIQSTTTT	.....VGVH	LVFDEQGRLL	TSTP.....T
3fiber	YVTLMGASDY	VNTLFKNKNV	.....SINVE	LYFDATGHIL	PDSSSLKTDL

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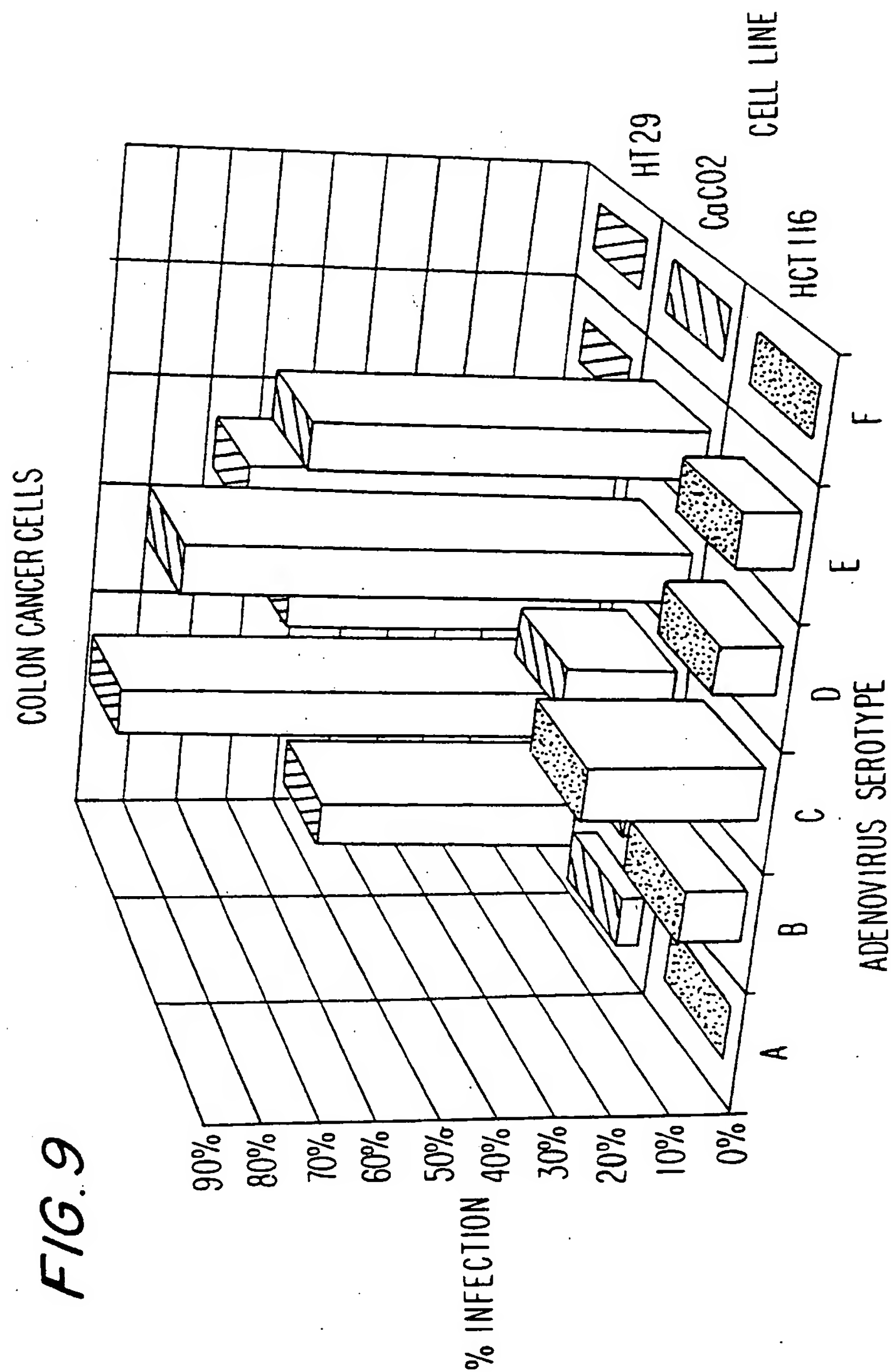
551	8fiber	..LGKSYWNF	RNQNSIMSTA	YEKAIGFMPN	LVAYPKPTTG	SKKY...ARD	600
	9fiber	..LGKSYWNF	RNENSIMSTA	YEKAIGFMPN	LVAYPKPTAG	SKKY...ARD	
	15fiber	..MDSSYWNY	RSDNSNLSQP	YKKAUGFMPS	KTAYPKQTKP	TNKEISQAKN	
	17fiber	..LDSTYWNF	RSDNLTVSEA	YKNAVEFMPN	LVAYPKPTTG	SKKY...ARD	
	2fiber	..LKKHYWNF	RNGNSTNANP	YTNAVGFMPN	LLAYPKTQSQ	T.....AKN	
	5fiber	..LDPEYWNF	RNGDLTEGTA	YTNAVGFMPN	LSAYPKSHGK	T.....AKS	
	4fiber	..TSKKYWGY	KQGDSIDGTP	YTNAVGFMPN	STAYPKTQSS	T.....TKN	
	40-1fiber	..LESSTWRY	QETNAVA...	..SNALTFMPN	STVYPRNKTA	D.....PGN	
	41fiber	..LESSTWRY	QETNAVA...	..SNALTFMPN	STVYPRNKTA	H.....PGN	
	40-2fiber	ILANSATWGY	RQGQSANTN.	VSNAVEFMPS	SKRYPNEKGS	E.....VQN	
	12fiber	ALVPQASWGY	RQGQSVSTNT	VTNGLGFMPN	VSAYPRPNAS	E.....AKS	
	3fiber	ELKYKQTADF	.....	..SARGFMPS	TTAYPFVLPN	AGTH...NEN	

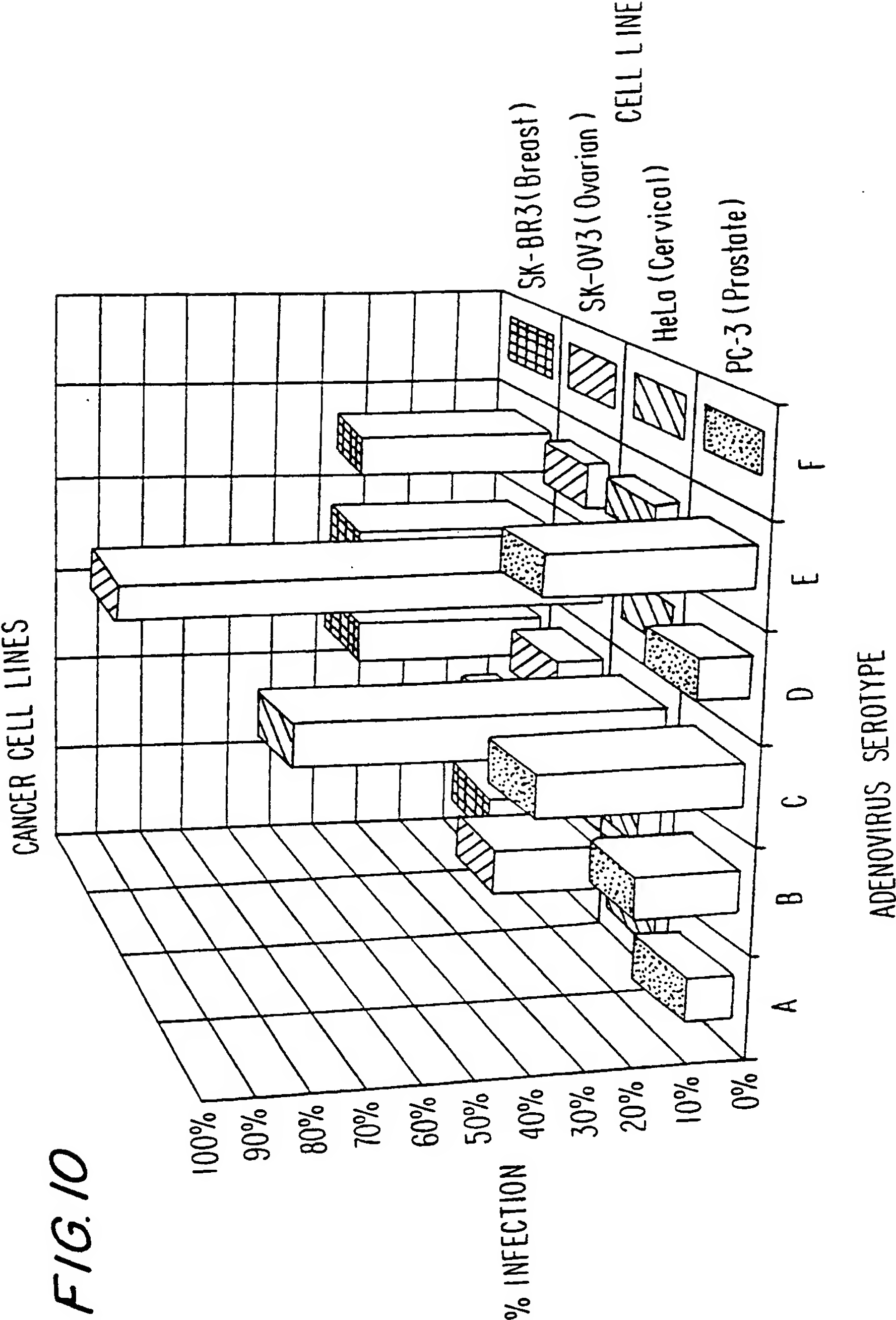
8fiber	601	IVYGNILYLG	KPHQ..	PVTI	KTFNQETG..	.....	CEYS	ITFDFSWAKT	650
9fiber		IVYGNILYLG	KPDQ..	PVTI	KTFNQETG..	.....	CEYS	ITFDFSWAKT	
15fiber		KIVSNVYLGG	KIDQ..	PCVI	IISFNEEAD..	.....	SDYS	IVFYFKWKYT	
17fiber		IVYGNILYLG	LAYQ..	PVVI	KVTFNEEAD..	.....	SAYS	ITFEFVWNKE	
2fiber		NIVSQVYLHG	DKTK..	PMIL	TITLNGTSES	TETSEVSTYS		MSFTWSWESG	
5fiber		NIVSQVYLN	DKTK..	PVTL	TITLNGTQET	GDTT.PSAYS		MSFSWDWSGH	
4fiber		NIVGQVYMNG	DVSK..	PMLL	TITLNGTDDT	T.....	SAYS	MSFSYTWING	
40-1fiber		MLI.....	QISP..	NITF	SVVYNEINS..	.....	GYA	FTFKW.SAEP	
41fiber		MLI.....	QISP..	NITF	SVVYNEINS..	.....	GYA	FTFKW.SAEP	
40-2fiber		MALTYTFLOQ	DPNM..	AISF	QSIYN..HA.	.....	IEGYS	LKFTW.RVRN	
12fiber		QMVSLTYLQ	DTSK..	PITM	KVAFNGITS..	.....	LNGYS	LTFMW.SGLS	
3fiber		YIFGQCYKA	SDGALFPLEV	TVMLNKRLPD		SRTSYVMTFL		WSLNAGLAPE	

FIG. 8D-2

8fiber	651	.YVNVEFETT	SFTFSYIAQE	..	672
9fiber		.YVNVEFETT	SFTFSYIAQE	..	
15fiber		.YENVQFDSS	SFNFSYIAQE	..	
17fiber		.YARVEFETT	SFTFSYIAQQ	..	
2fiber		KYTTETTFATN	SYTFSYIAQE	..	
5fiber		NYINEIFATS	SYTFSYIAQE	..	
4fiber		SYIGATFGAN	SYTFSYIAQQ	..	
40-1fiber		...GKPFHPP	TAVFCYITEQ	..	
41fiber		...GKPFHPP	TAVFCYITEQ	..	
40-2fiber		...NERFDIP	CCSFSYVTEQ	..	
12fiber		NYINQPFSTP	SCSFSYITQE	..	
3fiber		T.TQATLITS	PFTFSYIRED	D*	

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## INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 97/21494

A. CLASSIFICATION OF SUBJECT MATTER  
IPC 6 C12N15/86 A61K48/00

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C12N A61K C07K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	P.W. ROELVINK ET AL.: "Comparative analysis of adenovirus fiber-cell interaction: Ad2 and Ad9 utilize the same cellular fiber receptor but use different binding strategies for attachment" JOURNAL OF VIROLOGY, vol. 70, no. 11, November 1996, AMERICAN SOCIETY FOR MICROBIOLOGY US, pages 7614-7621, XP002062100 see page 7620, last paragraph	1-13
A	WO 96 26281 A (GENVEC INC ; CORNELL RES FOUNDATION INC (US)) 29 August 1996 see example 7	1,4,6-8, 10,11
	---	
	-/--	

☒ Further documents are listed in the continuation of box C.☒ Patent family members are listed in annex.

## \* Special categories of cited documents:

- \*A\* document defining the general state of the art which is not considered to be of particular relevance
- \*E\* earlier document but published on or after the international filing date
- \*L\* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- \*O\* document referring to an oral disclosure, use, exhibition or other means
- \*P\* document published prior to the international filing date but later than the priority date claimed

- \*T\* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- \*X\* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
- \*Y\* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.
- \*Z\* document member of the same patent family

Date of the actual completion of the international search

14 April 1998

Date of mailing of the international search report

123.04.98

Name and mailing address of the ISA

European Patent Office, P.B. 5818 Patentlaan 2  
NL - 2280 HV Rijswijk  
Tel. (+31-70) 340-2040, Tx. 31 651 epo nl  
Fax: (+31-70) 340-3016

Authorized officer

Cupido, M

# INTERNATIONAL SEARCH REPORT

International Application No  
PCT/US 97/21494

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		Relevant to claim No.
Category *	Citation of document, with indication, where appropriate, of the relevant passages	
A	J. GALL ET AL: "Adenovirus type 5 and 7 capsid chimera: Fiber replacement alters receptor tropism without affecting primary immune neutralization epitopes" JOURNAL OF VIROLOGY., vol. 70, no. 4, April 1996, pages 2116-2123, XP002050655 see the whole document	1,4,6-8, 10,11
P,X	--- WO 97 12986 A (CORNELL RES FOUNDATION INC) 10 April 1997 see page 15, line 1 - line 7 -----	1,2,13

# INTERNATIONAL SEARCH REPORT

Ir. .ational application No.

PCT/US 97/ 21494

## Box I Observations where certain claims were found unsearchable (Continuation of Item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☒ Claims Nos.: 11 to 13  
because they relate to subject matter not required to be searched by this Authority, namely:  
Although these claims are directed to a method of treatment of the human or animal body, the search has been carried out and based on the alleged effects of the adenoviral vector
2. ☐ Claims Nos.:  
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
3. ☐ Claims Nos.:  
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

## Box II Observations where unity of invention is lacking (Continuation of Item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
- ☐ No protest accompanied the payment of additional search fees.

# INTERNATIONAL SEARCH REPORT

information on patent family members

International Application No

PCT/US 97/21494

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO 9626281 A	29-08-96	AU 4980496 A CA 2213343 A EP 0811069 A	11-09-96 29-08-96 10-12-97
WO 9712986 A	10-04-97	NONE	